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(64) Human rhinovirus RNA and proteins.

(67) DNA fragments encoding all the structural and non-structural proteins and untranslated RNA genome regions of human rhinovirus type 14 have been molecularly cloned and their nucleotide sequence determined. The composition and sequence of the entire 7,212 nucleotide long genome RNA have been determined. Monoclonal antibodies have been produced which block the binding of virus to susceptible cells or which neutralize the infectivity of the virus.

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DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)
X	CHEMICAL ABSTRACTS, vol. 101, no. 19, 5th November 1984, page 142, abstract no. 164551k Columbus, Ohio, US; T. SKERN et al.: "Relationship of human rhinovirus strain 2 and poliovirus as indicated by comparison of the polymerase gene regions", & VIROLOGY, 1984, 136(1), 125-32 * Abstract *	6	C 12 N 15/00 C 12 N 5/00 C 12 P 21/00// (C 12 P 21/00: C 12 R 1:91)
P,X	CHEMICAL ABSTRACTS, vol. 103, no. 3, 22nd July 1985, page 270, abstract no. 19044j Columbus, Ohio, US; L.A. IVANOFF et al.: "Biochemistry and structure of picornavirus proteases", & PROG.CLIN.BIOL.RES. 1985, 180 (Intracell. Protein Catabolism) 255-6 * Abstract *	5	TECHNICAL FIELDS SEARCHED (Int. Cl. 4)
P,X	NUCLEIC ACIDS RESEARCH, vol. 12, no. 20, October 1984 pages 7859-7875 Cambridge, GB; G. STANWAY et al.: "The complete nucleotide sequence of a common cold virus: human rhinovirus 14" * Whole article *	1-6	C 12 N
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The present search report has been drawn up for all claims		-2-	
Place of search THE HAGUE		Date of completion of the search 02-05-1988	Examiner SKELLY
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document</p>			



European Patent
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CLAIMS INCURRING FEES

The present European patent application comprised at the time of filing more than ten claims.

- ☐ All claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for all claims.
- ☐ Only part of the claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claims:
- ☐ No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.

X LACK OF UNITY OF INVENTION

The Search Division considers that the present European patent application does not comply with the requirement of unity of invention and relates to several inventions or groups of inventions, namely:

1. Claims 1-6: DNA sequence corresponding to genomic RNA of human Rhinovirus type 14
2. Claims 7-9: Hybridoma cells and their monoclonal antibodies specific for human Rhinovirus proteins
3. Claims 10-20: Hybridoma cells and their monoclonal antibodies specific for cell receptors of human Rhinoviruses

- ☒ All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
- ☐ Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid, namely claims
- ☐ None of the further search fees has been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims:



DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)
	of human rhinovirus type 14"		
	* Whole article *	1-6	
Y	EP - A - 0 112 741 (INSTITUT PASTEUR)		
	* Whole document *	7-9	
Y	NATURE, vol. 259, 26th February 1976, pages 679-681 K. LONBERG-HOLM et al.: "The related animal viruses share receptors"		
	* Whole document *	7-20	
Y	ABS. ANNU. MEET. AM. SOC. MICROBIOL. vol. 82, 1982, abstract T 44 B.A. CAMPBELL et al.: "Production of monoclonal antibodies that inhibit attachment of Coxsackievirus B1 to HeLa cells"		
	* Abstract *	10-20	TECHNICAL FIELDS SEARCHED (Int. Cl. 4)
P, X	CHEMICAL ABSTRACTS, vol. 102, no. 7, 18th February 1985, page 449, abstract no. 60433c Columbus, Ohio, US; B. SHERRY et al.: "Evidence for at least two dominant neutralization antigens on human rhinovirus 14" & J. VIROL. 1985, 53(1), 137-43		
	* Abstract *	7-9	
P, Y	CHEMICAL ABSTRACTS, vol. 101, no. 11, 10th September 1984, page 332, abstract no. 87106q Columbus, Ohio, US; A. GORDON et al.: "Many rhinovirus serotypes share the same		
The present search report has been drawn up for all claims		-3-	
Place of search		Date of completion of the search	Examiner
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EUROPEAN SEARCH REPORT

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Application number
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-3-

DOCUMENTS CONSIDERED TO BE RELEVANT															
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)												
E	cellular receptor", & J.VIROL. 1984, 51 (2), 340-5 * Abstract *	7-20													
	US - A - 4 683 295 (CARSON) * Claims *	10-20													
			TECHNICAL FIELDS SEARCHED (Int. Cl. 4)												
The present search report has been drawn up for all claims															
Place of search		Date of completion of the search	Examiner												
<table border="0"><tr><td>CATEGORY OF CITED DOCUMENTS</td><td>T : the primary principle underlying the invention</td></tr><tr><td>X : particularly relevant if taken alone</td><td>E : earlier patent document, but published on, or after the filing date</td></tr><tr><td>Y : particularly relevant if combined with another document of the same category</td><td>D : document cited in the application</td></tr><tr><td>A : technological background</td><td>L : document cited for other reasons</td></tr><tr><td>O : non-written disclosure</td><td>& : member of the same patent family, corresponding document</td></tr><tr><td>P : intermediate document</td><td></td></tr></table>				CATEGORY OF CITED DOCUMENTS	T : the primary principle underlying the invention	X : particularly relevant if taken alone	E : earlier patent document, but published on, or after the filing date	Y : particularly relevant if combined with another document of the same category	D : document cited in the application	A : technological background	L : document cited for other reasons	O : non-written disclosure	& : member of the same patent family, corresponding document	P : intermediate document	
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P.O. Box 2000, Rahway New Jersey 07065 (US)**(43) Date of publication of application: 22.01.86
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LI LU NL SE**(74) Representative: **Warcoln, Jacques et al, Cabinet
Régimbeau 26, avenue Kléber, F-75116 Paris (FR)**(54) **Human rhinovirus RNA and proteins.**

(57) DNA fragments encoding all the structural and non-structural proteins and untranslated RNA genome regions of human rhinovirus type 14 have been molecularly cloned and their nucleotide sequence determined. The composition and sequence of the entire 7,212 nucleotide long genome RNA have been determined. Monoclonal antibodies have been produced which block the binding of virus to susceptible cells or which neutralize the infectivity of the virus.

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HUMAN RHINOVIRUS RNA AND PROTEINS

5

BACKGROUND OF THE INVENTION

Human rhinoviruses (HRV) belong to the
10 picornavirus family and contain at least 115
antigenically distinct serotypes. The picornavirus
family also includes poliovirus and hepatitis A
virus. Rhinoviruses are the most important common
cold viruses to be discovered. The name "rhinovirus"
15 reflects the prominent nasal involvement seen in
infections with these viruses. The number of
antigenically distinct rhinoviruses is so large that

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a person can be infected with a different rhinovirus each year and still not experience all of the known types in a life time.

HRV exist as a capsid structure (20-40 nm) with cubic symmetry utilizing 60 identical subunits composed of four peptide chains. The virions are ether resistant and acid labile. HRV contains an infectious, single-stranded genomic RNA approximately 7,200 bases in length. Upon entrance into a susceptible cell, the RNA is immediately translated into a 240,000 mw polyprotein using host cell translational machinery. This polyprotein is subsequently processed by both cellular and viral encoded proteases to yield the structural and non-structural proteins needed for completion of the infectious cycle.

Limited research has been performed on HRV because of the numerous serotypes involved and the poor growth and purification procedures for HRV used in the past. In addition, past procedures utilized in direct RNA and protein sequencing were very laborious and time consuming. However, recent technological advances in cloning and sequencing of nucleic acid has allowed molecular characterization of HRV. Using these techniques, the entire genomic RNA and corresponding amino acid sequence of a HRV could be determined. This information would define significant target areas of the virus (i.e., proteolytic cleavage sites) for which specific antiviral agents could be developed. It has also enabled the isolation of DNA fragments which could be used as probes to determine common genomic regions

among the many serotypes of rhinoviruses. This would allow construction of a genomic map of common and unique sequences within the rhinovirus family.

5 OBJECTS OF THE INVENTION

It is an object of the present invention to provide a method for isolating and cloning of DNA representative of the genomic RNA of human rhinoviruses. Another object is to determine the
10 nucleotide and derived amino acid sequences of HRV type 14 genomic RNA. A third object is to determine the amino acid sequence of precursor proteins utilized during polyprotein processing. Yet another object is to provide a method for producing HRV
15 proteins or subunits thereof by expression of cloned DNA in an appropriate host. Another object is to provide HRV proteins or subunits thereof which will be useful as immunogens to raise antibodies to HRV. Still another object is to provide a vector
20 containing DNA representative of HRV type 14 genomic RNA or part thereof. A further object is to provide for construction of an infectious cDNA plasmid capable of yielding infectious virus particles when placed in an appropriate host.
25 Another object is to produce a monoclonal antibody capable of blocking the binding of a major group of rhinoviruses. A further object is to produce monoclonal antibodies to rhinovirus structural proteins (VP1, VP2 and VP3) that neutralize the
30 infectivity of the virus. These and other objects of the present invention will be apparent from the following description.

SUMMARY OF THE INVENTION

DNA fragments encoding the entire 9212 nucleotides of HRV 14 genome RNA have been
5 molecularly cloned and their nucleotide sequence completely determined. Eight monoclonal antibodies have been isolated which neutralize infections HRV-14 virions. In addition, a mouse monoclonal antibody was isolated which specifically blocked the
10 attachment of at least 75 antigenically distinct serotypes of HRVs to cellular receptors.

DETAILED DESCRIPTION

Plaque purified human rhinovirus type 14 was
15 grown and purified by isopycnic banding in a metrizamide (2-[3-acetamido-5-N-methylacetamido-2,4,6-triiodobenzamido]-2-deoxy-D-glucose) gradient. Purified virus was disrupted with detergents and capsid proteins digested with proteinase K. Genomic
20 RNA was isolated by oligo (dT) cellulose chromatography and purified by velocity sedimentation in a sucrose gradient. RNA sedimenting at 35S, relative to ribosomal RNA markers, was collected and ethanol precipitated.

25 Recovered genomic RNA was used as a template for synthesis of its complementary strand to produce a double stranded DNA molecule corresponding to at least a portion of the original genome RNA. The double stranded DNA was modified to provide "sticky
30 ends" and was placed into a bacterial plasmid pBR 322. Prokaryotic organisms were exposed to the resulting vector and those which stably incorporated

the vector were identified and isolated.

Vector DNA was extracted from these hosts and this material was characterized. Vector DNA was first analyzed for size of the HRV cDNA insert and
5 vectors containing the largest inserts mapped for their relationship to the viral RNA genome and each other. Selected vectors containing DNA sequences representing the entire length of the HRV genome RNA were sequenced using a known sequencing method.
10 Fragments of these vectors were radioactively labeled and isolated prior to sequencing. By overlapping the sequences obtained from these fragments, the entire HRV 14 sequence was determined. Subsequent conversion of the DNA sequence to RNA and protein was
15 made by computer. Determination of the amino acid sequence allows one to determine the structure of the virus and to identify its structural and non-structural proteins. Knowledge of the viral structure allows inferences to be made as to likely
20 viral receptor sites, proteolytic cleavage sites, and neutralization sites of the virus. The cloned DNA is available for genetic analysis and modification and for use in the development of antiviral compounds. Expression in suitable hosts would yield large
25 quantities of viral proteins for development of antiviral compounds or subunit vaccines.

The use of recombinant DNA techniques is described in many published articles, for example, Maniatis et al, Molecular Cloning, A Laboratory
30 Manual, Cold Spring Harbor, New York, 1982, the disclosure of which is hereby incorporated by reference.

In order to generate monoclonal antibodies that neutralize the infectivity of HRV 14 or that block the binding of many HRV serotypes to the virus receptor on susceptible cells, mice were immunized with either purified virions or whole cells and cell membranes enriched for HRV receptor sites, e.g., HeLa cells. Hybridomas were produced following fusion of spleen cells of immunized mice with mouse myeloma cells. From the resulting hybridoma cell lines eight monoclonal antibodies were isolated which are capable of neutralizing HRV-14. In addition, a monoclonal antibody was selected that was capable of blocking the attachment of at least 75 different serotypes of HRV to receptor sites on susceptible cells. The specific serotypes which were unable to cause infection in the presence of the monoclonal antibody as well as those serotypes which were able to cause infection in the presence of the monoclonal antibody of the present invention are shown in Table I.

As a result of these experiments it has been determined that human rhinoviruses consist of a major group and a minor group. The major group is neutralized or rendered incapable of causing infection by the monoclonal antibody while the minor groups seems unaffected by the monoclonal antibody.

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TABLE I

5

CHARACTERIZATION OF MONOCLONAL ANTIBODY
TO HRV CELLULAR RECEPTOR

ANTIBODY TYPE: IgG-1

		<u>PROTECTION AGAINST INFECTION BY:</u>							
10	HRV-3	HRV-12	HRV-21	HRV-33	HRV-42	HRV-57	HRV-67	HRV-77	
	HRV-4	HRV-13	HRV-22	HRV-34	HRV-46	HRV-58	HRV-68	HRV-7E	
	HRV-5	HRV-14	HRV-23	HRV-35	HRV-48	HRV-59	HRV-69	HRV-7F	
	HRV-6	HRV-15	HRV-24	HRV-36	HRV-50	HRV-60	HRV-70	HRV-80	
	HRV-7	HRV-16	HRV-25	HRV-37	HRV-51	HRV-61	HRV-71	HRV-81	
	HRV-8	HRV-17	HRV-26	HRV-38	HRV-52	HRV-63	HRV-72	HRV-83	
	HRV-9	HRV-18	HRV-27	HRV-39	HRV-54	HRV-64	HRV-73	HRV-84	
	HRV-10	HRV-19	HRV-28	HRV-40	HRV-55	HRV-65	HRV-74	HRV-85	
15	HRV-11	HRV-20	HRV-32	HRV-41	HRV-56	HRV-66	HRV-75	HRV-86	
							HRV-76	HRV-8E	
							HRV-HANKS		

20

NO PROTECTION AGAINST INFECTION BY:

HRV-1A	HRV-29	HRV-44	HRV-62
HRV-1B	HRV-30	HRV-47	HRV-87
HRV-2	HRV-31	HRV-49	

25

30

It has also been found that the monoclonal antibody of the present invention binds strongly to susceptible cells and possesses greater avidity for the cell as it actually displaces the major group of human rhinoviruses bound to the cell. This characteristic of the antibody of the present invention is shown in Table II. Human rhinoviruses labeled with S³⁵ methionine were bound to susceptible cell membranes in vitro and unbound virus was removed by washing.

Increasing amounts of the monoclonal antibody of the present invention were added to the membranes and the amount of bound virus that was released was measured.

The monoclonal antibody of the present invention is useful to prevent HRV infection or to ameliorate the duration and severity of infection. Treatment of susceptible cells with the monoclonal antibody of the present invention renders the cells resistant to viral infection for a period of up to 12 hours following removal of excess antibody. The monoclonal antibody may be prepared in a suitable topical formulation for administration in the form of drops or as a spray. The dosage may range of from about 1.5 to about 150 µg given at intervals of from about 2 hours to about 12 hours, and more usually in a range of from about 5 to about 25 µg given at intervals of about 4, 8 or 12 hours. A typical formulation employs the monoclonal antibody of the present invention in PBS containing a stabilizer, e.g., 20% SPGA.

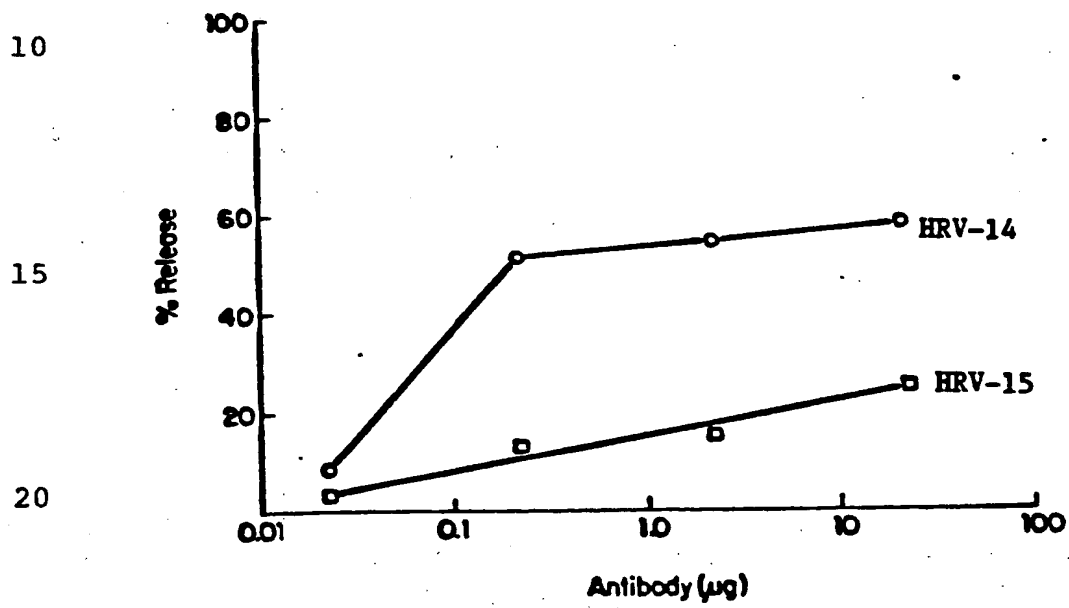
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TABLE II

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25

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The following examples illustrate the present invention without, however, limiting the same thereto. The disclosure of each reference mentioned in the following examples is hereby incorporated by
5 reference.

EXAMPLE 1

Virus Growth and Purification

10 HRV 14 was obtained from the American Type Culture collection (Rockville, MD), ATCC VR-284 and plaque purified by standard techniques (Yin et al., J. Virol. 12:108-113, 1973). Plaque purified HRV was propagated on HeLa R19 cells, a subclone of HeLa
15 cells derived by standard procedures (Coller et al., Hybridoma 2:91-96, 1983). Cells were routinely propagated in growth medium (GM): McCoy's 5A medium containing 10% fetal calf serum, 30 mM MgCl₂ and 20 units/ml penicillin and 20 µg/ml streptomycin.
20 HeLa R19 monolayers were infected at a MOI of 1 and harvested when cells detached freely from the flask surface (usually 16-24 hours). The cells suspension was frozen and quickly thawed to release virus particles from the cells. After clarification by
25 centrifugation at slow speed (4000 x g for 5 minutes at 4°C), Polyethylene glycol 6000 (PEG-6000) and NaCl were added to concentrations of 7% and 2.2% respectively and the mixture stirred at 4°C for 4-16 hours. The precipitated virus was recovered by
30 centrifugation (10,500 x g for 15 minutes at 4°C) and resuspended in 10 mM R-buffer (a solution containing 10 mM tris-HCl pH 7.5, 1 mM EDTA [ethylenediamine

tetraacetic acid], 0.2 M NaCl, 50 mM MgCl₂ and 10% glycerol). Sodium deoxycholate and polyoxyethylene (9) octaphenol were added to concentrations of 0.3% and 0.6%, respectively, for 30 minutes on ice and the suspension clarified by centrifugation (4000 x g for 5 minutes at 4°C). The supernatant liquid (7.5 ml) was layered over a 5 ml linear density gradient of 40-60% (w/v) metrizamide in R-buffer lacking the glycerol. Isopycnic banding of the virus sample was achieved by centrifugation for 24 hours at 150,000 x g in a Beckman SW40 rotor. Virus bands were generally the densest of the visible bands in the gradient and were harvested manually by bottom puncture. After dilution, 3-fold in R-buffer the virus was repelleted by centrifugation at 200,000 x g for 2 hours.

EXAMPLE 2

Genomic RNA Isolation

HRV 14 purified from forty 150 cm² flasks of HeLa R19 cells was resuspended in 0.9 ml 10 mM Tris-HCl, 1 mM EDTA, 0.3% SDS (sodium dodecyl sulfate), pH 7.5. Proteinase K was added to a final concentration of 0.5 mg/ml and the viral proteins digested for 30 minutes at 37°C. The digested mixture was heated to 95°C for 1 minute and then quick chilled in an ice bath. NaCl was added to 0.5 M and the solution passed through a 1 cm x 0.7 cm column of oligo (dT) cellulose column (Collaborative Research, Lexington, Mass.) Genomic RNA retained on the column was eluted with H₂O and precipitated at

42°C for 2 hours and then stopped by the addition of 5 µl of 0.5M EDTA and 5 µl of 5M NaOH. Following an additional 30 minutes at 37°C, the synthesized cDNA was isolated by cellulose (Whatman CF-11) column chromatography (Franklin, P.N.A.S. 55: 565, 1966) and ethanol precipitated at -20°C.

The cDNA was pelleted and resuspended in 40 µl of H₂O and 50 µl of a solution containing 0.2M Hepes pH 6.9, 20 mM MgCl₂, 5 mM DTT, 0.14M KCl, and 1 mM each of dATP, dCTP, dGTP, dTTP. The second DNA strand was then synthesized by the addition of 10 µl of the large fragment of DNA polymerase I, 50 units (New England Biolabs). The reaction mix was incubated for 16 hours at 15°C and stopped by the addition of 2 µl of 0.5M EDTA. The double stranded DNA was phenol/chloroform extracted and precipitated with ethanol. To aid in the completion of the second strand synthesis, the double stranded DNA was resuspended in 30 µl of H₂O, 5 µl of 1M Tris pH 8.3, 7 µl of 1M KCl, 2 µl of 250 mM MgCl₂, 2.5 µl each of 20 mM dCTP, dGTP, dTTP and 10 mM dATP, 2 µl of 2-mercaptoethanol, and 2 µl (30 units) of reverse transcriptase. The reaction was incubated at 42°C for 1 hour and stopped by the addition of 2 µl of 0.5M EDTA. The double stranded cDNA was ethanol precipitated at -20°C.

EXAMPLE 4Conversion of cDNA "Hairpin Structure" to Linear Form

The order to insert cDNA into plasmid
5 vectors for cloning, it is necessary to cleave the
"hairpin structure" formed during synthesis into a
linear form possessing accessible termini at each end
of the molecule. This was carried out as follows;
the cDNA, previously synthesized in Example 3, was
10 resuspended in 79 μ l H_2O , and 20 μ l of a solution
containing 1M NaCl, 0.25M NaAcetate pH 4.5, 5 mM
 $ZnSO_4$ and 2.5% v/v glycerol. S_1 nuclease, 290
units, (PL Biochemicals, Piscataway, N.J.) was added
and the reaction incubated 30 minutes at 37°C. The
15 reaction was stopped by the addition of 2 μ l of 0.5M
EDTA, phenol/chloroform extracted and the DNA
purified by chromatography on a cellulose column
(Whatman CF-11) as described by Franklin, supra.

20

EXAMPLE 5Addition of Oligo dC to 3' Ends of cDNA

To allow insertion of cDNA into oligo
deoxyguanosine (dG) "tailed" pBR322, oligo
25 deoxycytidine (dC) "tails" were added to the cDNA
from Example 4. Following ethanol precipitation, the
cDNA (approximately 2 μ g) was resuspended in 149 μ l
of H_2O , 40 μ l of 1M potassium cacodylate buffer pH
7.0, 4 μ l of 0.1M $CoCl_2$, 2 μ l of 0.1M 2-mercapto-
30 ethanol, 1 μ l of 0.1 M [3H] dCTP (150 μ Ci) and 4 μ l
(60 units) terminal deoxytransferase (PL
Biochemicals). The reaction was incubated at 37°C

and aliquots removed at 1 minute (80 μ l), 2 minutes (80 μ l), and 3 minutes (40 μ l) and pooled together on ice. The "tailed" cDNA was ethanol precipitated at -20°C.

5

EXAMPLE 6Vector/cDNA Annealing and Cell Transformation

To insert the tailed cDNA from Example 5
10 into pBR322, 25 ng of pBR322 oligo dG cloning vector (BRL, Bethesda, MD) was annealed with 13, 26, and 39 ng of oligo dC tailed cDNA in 40 μ l of a solution containing 10 mM Tris-HCl pH 7.5, 1 mM EDTA and 0.1M NaCl at 65°C for 5 minutes and 56°C for 90 minutes.
15 After annealing, each reaction mix was cooled to 0°C and added to 200 μ l calcium treated E. coli strain RRI (Maniatis et al., op. cit., p. 250). After a 30 minute incubation on ice, the reaction was heated for 2 minutes at 42°C, 1 ml of L broth (Bacto tryptone 10 g, Bacto yeast extract 5 g, and NaCl 5 g, brought up
20 to 1 l with distilled water) added, and the transformation reaction incubated at 37° for 30 minutes. Each transformation reaction was plated (250 μ g) onto 4 agar petri plates containing
25 tetracycline (15 μ g/ml) (Maniatis et al., op. cit., p. 70-72). Each plate yielded approximately 30 colonies after 16 hours of incubation at 37°C.

EXAMPLE 7Characterization of Selected Clones

Colonies (354) from Example 6 were picked
5 with a toothpick and used to inoculate 200 μ l of L
broth containing 12 μ g/ml tetracycline in 96 well
serological plates. Following a 16 hour incubation
at 37°C, 20 μ l of the new growth culture was used to
inoculate 2 ml L broth containing 12 ng Na_2HPO_4 ,
10 6 ng KH_2PO_4 , 1 ng NaCl, 2 ng NH_4Cl , and 4 μ g
glucose. After shaking at 37°C for 16 hours, 0.5 ml
was frozen after adding glycerol to 15% and stored at
-70°C. The remaining 1.5 ml was pelleted 1 minute at
12,000 x g and the supernatant liquid discarded. The
15 pellet was resuspended in 0.15 ml of a solution
containing 8% w/v sucrose, 50 mM EDTA, 50 mM Tris-HCl
pH 8.0, 5% v/v polyoxyethylene (9) octaphenol and 0.6
mg/ml lysozyme and incubated 5 minutes at 25°C. The
bacteria were then placed in a boiling water bath for
20 40 seconds and immediately plunged into an ice bath.
Following centrifugation for 15 minutes at 12,000 x
g, the pellet was removed with a toothpick and
discarded. An equal volume of isopropanol was added
to the supernatant liquid and the solution mixed and
25 placed at -20°C for 10 minutes. After pelleting for
5 minutes at 12,000 x g, the supernatant liquid was
removed and the pellet washed with 70% v/v ethanol
and dried. The resulting plasmid DNA was resuspended
in 50 μ l H_2O and stored at -20°C.
30 To determine the size of each cDNA insert,
10 μ l of each plasmid preparation was digested with
the restriction endonuclease Pst I (Boehringer

Mannheim, Indianapolis IN) as described by the manufacturer. The digests were analyzed by electrophoresis in a 1% agarose gel as described (Maniatis et al., op. cit., p. 150). Three cDNA
5 clones, designated 7, 57, and 186, were selected based on size (2000 bp) and colony hybridization studies (Grunstein and Hogness, Proc. Nat. Acad. Sci. U.S.A. 72:3691-3965, 1975) which indicated that 7
10 (3200 bp) and 186 (3200 bp) did not overlap and that 57 (2000 bp) bridged both 7 and 186.

EXAMPLE 8

Construction of Deletion Subclones

15 Twenty micrograms of clones 7 and 186 (Example 7) were digested with 20 units of restriction endonuclease Pst I in a total volume of 30 μ l as described by the manufacturer. After
20 separation of plasmid DNA from viral cDNA on a 1% low melt agarose gel (FMC Corporation, Rockland, ME), each cDNA insert was extracted by the method of Tautz and Renz (Anal. Biochem. 132:14-19, 1983) 20 μ g
bacterial plasmid pUC9 DNA (Vieiera et al., Gene 19: 259-268, 1982) was digested with Pst I as described
25 above and mixed with 30 μ l of a 1M solution of Tris-HCl pH 8.0 containing 20 units of alkaline phosphatase (Boehringer Mannheim) for 2 hours at 65°C. The mixture was extracted twice with
phenol/chloroform and the pUC9 DNA ethanol
30 precipitated. One μ g (0.5 p mole) of both insolated clone 7 and 186 inserts were ligated separately to 2.8 μ g (1.0 p mole) of digested pUC9 DNA with 2.5

units of T_4 DNA ligase (BRL, Gaithersburg, MD) in 10 μ l of a solution containing 5 mM Tris-HCl pH 8.0, 10 mM $MgCl_2$, 20 mM DTT, 1 mM ATP, and 0.001% polyoxyethylene (9) octaphenol at 15°C for 2 hours.

- 5 E. coli strain HB101 (Gene 6:23-28, 1979) was transformed with each of the above ligation mixtures and transformants selected on agar plates containing 100 μ g/ml ampicillin. The orientation of each insert in pUC9 plasmid was determined in
- 10 relation to the orientation of the LAC gene. An "A" orientation of the insert is the same orientation as the LAC gene and a "B" is the reverse. Two hundred μ g of pUC9 (7A) and pUC9 (186B, were linearized with
- 15 DNase I (Boehringer Mannheim) in the presence of 1 mM $MnCl_2$ and DNAs precipitated with PEG 6000 according to the method of Hong (J. Mol. Bio. 158:539-549, 1982). Full length DNA was separated by gel
- 20 electrophoresis (described above) and recovered from the agarose by the method of Wieslander (Anal. Biochem. 98:305, 1979). Ten μ g of linearized pUC9 (7A) or pUC9 (186B) were digested with 20 units of restriction endonuclease Sal I (Boehringer Mannheim) in 40 μ l as described by the manufacturer. The
- 25 reaction mixtures were then heated to 65°C for 5 minutes and 3 μ l 1.0M Tris-HCl pH 7.5, 1.5 μ l 0.25M $MgCl_2$, 12 μ l of a solution containing 0.1 mM each of dATP, dCTP, dGTP, and dTTP, 2.5 μ l H_2O , and 1 μ l (2.5 units) "large fragment" of DNA polymerase
- 30 (Boehringer Mannheim) were added and incubated at 37°C for 30 minutes. Following an incubation at 65°C for 10 minutes, the DNA was precipitated by adding 60 μ l of 10% PEG-6000 and 1.25M NaCl and incubated on

ice for 1.5 hours. The DNA was collected by centrifugation (12,000 x g) for 10 minutes and dissolved in 50 μ l of a solution containing 10 mM Tris-HCl pH 7.5, 1 mM EDTA and 10 mM NaCl. The DNA
5 was then extracted by phenol/chloroform and ethanol precipitated for 10 minutes at -70°C. The final precipitate was dissolved in 10 μ l of a solution containing 10 mM Tris-HCl pH 7.5, 1 mM EDTA and 10 mM NaCl, and mixed with 1.1 μ l of a solution
10 containing 0.5M Tris-HCl pH 8.0, 0.1M $MgCl_2$, 0.2M DTT, 0.01M ATP, and 0.01% polyoxyethylene (9) octaphenol, and 1 μ l (10 units) T_4 DNA ligase (New England Nuclear). The mixture was incubated at 15°C for 4 hours and used to transform E. coli strain
15 HB101 cells as described above. From 10 μ g of pUC9 (7A) and pUC9 (186B), 10,000 and 17,000 transformants were obtained, respectively.

Deletion clones (transformants) were screened as described in Example 7 and the size of
20 each DNA insert was determined by restriction endonuclease digestion with BAM HI [pUC9 (186B) clones] and with Pst I and Bam HI [pUC9 (7A) clones].

EXAMPLE 9

25 Nucleic Acid Sequencing of cDNA Inserts

Digestion of clones 7, 57, and 186 and their subset of deletion clones (Example 8) with various commercially available restriction enzymes generated
30 a subset of overlapping DNA fragments. These fragments were radioactively labeled one of two ways.

When restriction enzyme digestion resulted in a 5' overhang in double stranded DNA (i.e., XBA I), 25 µg of the DNA were labeled by resuspending in 80 µl of a solution containing 50 mM NaCl, 6 mM Tris-HCl pH 7.5, 6 mM MgCl₂, 6 mM 2-mercaptoethanol and 150-200 uCi [α^{32} P] deoxynucleotide triphosphate representing the first nucleotide to be specified by the overhang, and 5 units of "large fragment" of DNA polymerase I (Boehringer Mannheim) and incubated at 15°C for 2 hours. Alternatively, when the restriction enzyme digestion resulted in a 3' overhang in the DNA (i.e. Pst I), the DNA was labeled at the 3' end by resuspension in 40 µl of a solution containing 0.2M K cacodylic acid pH 7.0, 2 mM CaCl₂, 2 mM 2-mercaptoethanol, 250 uCi [α^{32} P] dd ATP (Amersham, Chicago, IL) and 20 units terminal deoxytransferase (PL Biochemicals) and incubated 1 hour at 37°C.

Both reactions were stopped by the addition of 4 µl of 0.5M EDTA and labeled DNA separated from unincorporated triphosphates by chromatography through an "Elutip" (Schleicher and Schuell, Keene, NH) using the supplier's protocol. Purified DNA was precipitated with ethanol and recut with a second restriction endonuclease which cut asymmetrically between the radioactively labeled ends, generating at least 2 fragments for each labeled fragment. The cut fragments were separated by electrophoresis in a 5% (w/v) acrylamide gel [15 x 15 x 0.015 cm] containing 42 ml of 29% w/v acrylamide (Bio Rad), 1% w/v N,N'-bis-acrylylcystamine (Bio Rad), 0.1% w/v ammonium persulfate, 0.38% N,N,N',N'-tetramethylene-

diamine (Bio Rad), 90 mM Tris base, 140 mM boric acid, and 1 mM EDTA] at 20 mA for 3 hours. The gel was stained in 200 ml of a solution containing 10 mM Tris-HCl pH 7.5, 1 mM EDTA and 1 µg/ml ethidium bromide for 30 minutes and DNA bands visualized by UV light. Appropriate bands were excised and the DNA recovered by dissolving the acrylamide gel slice in 1 ml of a solution containing 0.2M NaCl, 20 mM Tris pH 7.4, 1 mM EDTA and 40 mM 2-mercaptoethanol for 15 minutes at 25°C. The dissolved gel solution was then chromatographed through an "Elutip" (see above) and recovered DNA ethanol precipitated.

EXAMPLE 10

15

Sequence Determination

DNA fragments, labeled with [³²P] at only one end as described in Example 9, were sequenced using the chemical DNA sequencing methods described by Maxam and Gilbert (Methods in Enzymology, Vol. 65. 1980). Synthetic deoxynucleotide primers and Sanger (Zimmern et al., P.N.A.S. 75: 4257-4261, 1978) dideoxy sequencing techniques were used to complete the determination of the nucleotide sequence. A continuous DNA sequence of 7,209 nucleotides was obtained using the Intelligenetics software package. This nucleotide sequence and the amino acids inferred therefrom follow.

30

TTAAAACAGCG GATGGGTATC CCACCATTCG ACCCATTTGGG TGTAGTACTC
TGGTACTATG TACCTTTGTA CGCCTGTTTC TCCCCAACCA CCCTTCCTTA
AAATTCCCAC CCATGAAACG TTAGAAGCTT GACATTAAAG TACAATAGGT
5 GCGGCCATAT CCAATGGTGT CTATGTACAA GCACTTCTGT TTCCCAGGAG
CGAGGTATAG GCTGTACCCA CTGCCAAAAG CCTTTAACCG TTATCCGCCAA
CCAACTACGT AACAGTTAGT ACCATCTTGT TCTTGACTGG ACGTTCGATC
AGGTGGATTT TCCCTCCACT AGTTTGGTCG ATGAGGCTAG GAATTCCCCA
CGGGTGACCG TGTCCTAGCC TGCCTGGCGG CCAACCCAG CTTATGCTGG
10 GACGCCCTTT TAAGGACATG GTGTGAAGAC TCGCATGTGC TTGGTTGTGA
GTCCTCCGGC CCCTGAATGC GGCTAACCTT AACCTAGAG CCTTATGCCA
CGATCCAGTG GTTGTAAGGT CGTAATGAGC AATTCCGGGA CGGGACCGAC
TACTTTGGGT GTCCGTGTTT CTCATTTTTC TTCATATTGT CTTATGGTCA
CAGCATATAT ATACATATAC TGTGATC

15

VP4

ATG GGC GCT CAG GTT TCT ACA CAG AAA AGT GGA TCT CAC
MET Gly Ala Gln Val Ser Thr Gln Lys Ser Gly Ser His

20 GAA AAT CAA AAC ATT TTG ACC AAT GGA TCA AAT CAG ACT
Glu Asn Gln Asn Ile Leu Thr Asn Gly Ser Asn Gln Thr

TTC ACA GTT ATA AAT TAC TAT AAG GAT GCA GCA AGT ACA
Phe Thr Val Ile Asn Tyr Tyr Lys Asp Ala Ala Ser Thr

25

TCA TCA GCT GGT CAA TCA CTG TCA ATG GAC CCA TCT AAG
Ser Ser Ala Gly Gln Ser Leu Ser MET Asp Pro Ser Lys

30 TTT ACA GAA CCA GTT AAA GAT CTC ATG CTT AAG GGT GCA
Phe Thr Glu Pro Val Lys Asp Leu MET Leu Lys Gly Ala

VP4 VP2

CCA GCA TTG ATT TCA CCC AAT GTT GAG GCC TGT GGT TAT
Pro Ala Leu Asn Ser Pro Asn Val Glu Ala Cys Gly Tyr

5 AGT GAT AGA GTA CAA CAA ATC ACA CTC GGG AAT TCA ACA
Ser Asp Arg Val Gln Gln Ile Thr Leu Gly Asn Ser Thr

ATA ACA ACA CAA GAA GCA GCC AAC GCT GTT GTG TGT TAT
Ile Thr Thr Gln Glu Ala Ala Asn Ala Val Val Cys Tyr

10 GCT GAA TGG CCA GAG TAC CTT CCA GAT GTG GAC GCT AGT
Ala Glu Trp Pro Glu Tyr Leu Pro Asp Val Asp Ala Ser

GAT GTC AAT AAA ACT TCA AAA CCA GAC ACT TCT GTC TGT
15 Asp Val Asn Lys Thr Ser Lys Pro Asp Thr Ser Val Cys

AGG TTT TAC ACA TTG GAT AGT AAG ACA TGG ACA ACA GGT
Arg Phe Tyr Thr Leu Asp Ser Lys Thr Trp Thr Thr Gly

20 TCT AAA GGC TGG TGC TGG AAA TTA CCA GAT GCA CTC AAG
Ser Lys Gly Trp Cys Trp Lys Leu Pro Asp Ala Leu Lys

GAT ATG GGT GTG TTC GGG CAA AAC ATG TTT TTC CAC TCA
Asp MET Gly Val Phe Gly Gln Asn MET Phe Phe His Ser

25 CTA GGA AGA TCA GGT TAC ACA GTA CAC GTT CAG TGC AAT
Leu Gly Arg Ser Gly Tyr Thr Val His Val Gln Cys Asn

GCC ACA AAA TTC CAT AGC GGT TGT CTA CTT GTA GTT GTA
30 Ala Thr Lys Phe His Ser Gly Cys Leu Leu Val Val Val

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17112IA

ATA CCA GAA CAC CAA CTG GCT TCA CAT GAG GGT GGC AAT
Ile Pro Glu His Gln Leu Ala Ser His Glu Gly Gly Asn

5 GTT TCA GTT AAA TAC ACA TTC ACG CAT CCA GGT GAA CGT
Val Ser Val Lys Tyr Thr Phe Thr His Pro Gly Glu Arg

GGT ATA GAT TTA TCA TCT GCA AAT GAA GTG GGA GGG CCT
Gly Ile Asp Leu Ser Ser Ala Asn Glu Val Gly Gly Pro

10 GTC AAG GAT GTC ATA TAC AAT ATG AAT GGT ACT TTA TTA
Val Lys Asp Val Ile Tyr Asn MET Asn Gly Thr Leu Leu

GGA AAT CTG CTC ATT TTC CCT CAC CAG TTC ATT AAT CTA
Gly Asn Leu Leu Ile Phe Pro His Gln Phe Ile Asn Leu

15 AGA ACC AAT AAT ACA GCC ACA ATA GTG ATA CCA TAC ATA
Arg Thr Asn Asn Thr Ala Thr Ile Val Ile Pro Tyr Ile

AAC TCA GTA CCC ATT GAT TCA ATG ACA CGT CAC AAC AAT
20 Asn Ser Val Pro Ile Asp Ser MET Thr Arg His Asn Asn

GTC TCA CTG ATG GTC ATC CCT ATT GCC CCT CTT ACA GTA
Val Ser Leu MET Val Ile Pro Ile Ala Pro Leu Thr Val

25 CCA ACT GGA GCA ACT CCC TCA CTC CCT ATA ACA GTC ACA
Pro Thr Gly Ala Thr Pro Ser Leu Pro Ile Thr Val Thr

ATA GCA CCT ATG TGC ACT GAG TTC TCT GGG ATA AGG TCC
Ile Ala Pro MET Cys Thr Glu Phe Ser Gly Ile Arg Ser

30

VP2 VP3

AAG TCA ATT GTG CCA CAA GGT TTG CCA ACT ACA ACT TTG
Lys Ser Ile Val Pro Gln Gly Leu Pro Thr Thr Thr Leu

5 CCG GGG TCA GGA CAA TTC TTG ACC ACA GAT GAC AGG CAA
Pro Gly Ser Gly Gln Phe Leu Thr Thr Asp Asp Arg Gln

TCC CCC AGT GCA CTG CCA AAT TAT GAG CCA ACT CCA AGA
Ser Pro Ser Ala Leu Pro Asn Tyr Glu Pro Thr Pro Arg

10 ATA CAC ATA CTA GGG AAA GTT CAT AAC TTG CTA GAA ATT
Ile His Ile Leu Gly Lys Val His Asn Leu Leu Glu Ile

ATA CAG GTA GAT ACA CTC ATT CCT ATG AAC AAC ACG CAT
15 Ile Gln Val Asp Thr Leu Ile Pro MET Asn Asn Thr His

ACA AAA GAT GAG GTT AAC AGT TAC CTC ATA CCA CTA AAT
Thr Lys Asp Glu Val Asn Ser Tyr Leu Ile Pro Leu Asn

20 GCA AAC AGG CAA AAT GAG CAG GTT TTT GGG ACA AAC CTG
Ala Asn Arg Gln Asn Glu Gln Val Phe Gly Thr Asn Leu

TTT ATT GGT GAT GGG GTC TTC AAA ACT ACT CTT CTG GGT
Phe Ile Gly Asp Gly Val Phe Lys Thr Thr Leu Leu Gly

25 GAA ATT GTT CAG TAC TAT ACA CAT TGG TCT GGA TCA CTT
Glu Ile Val Gln Tyr Tyr Thr His Trp Ser Gly Ser Leu

AGA TTC TCT TCG ATG TAT ACT GGT CCT GCC TTG TCC AGT
30 Arg Phe Ser Ser MET Tyr Thr Gly Pro Ala Leu Ser Ser

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GCT AAA CTC ACT CTA GCA TAC ACC CCG CCT GGT GCT CGT
Ala Lys Leu Thr Leu Ala Tyr Thr Pro Pro Gly Ala Arg

5 GGT CCA CAG GAC AGG AGA GAA GCA ATG CTA GGT ACT CAT
Gly Pro Gln Asp Arg Arg Glu Ala MET Leu Gly Thr His

GTT GTC TGG GAT ATT GGT CTG CAA TCC ACC ATA GTA ATG
Val Val Trp Asp Ile Gly Leu Gln Ser Thr Ile Val MET

10 ACA ATA CCA TGG ACA TCA GGG GTG CAG TTT AGA TAT ACT
Thr Ile Pro Trp Thr Ser Gly Val Gln Phe Arg Tyr Thr

GAT CCA GAT ACA TAC ACC AGT GCT GGC TTT CTA TCA TGT
Asp Pro Asp Thr Tyr Thr Ser Ala Gly Phe Leu Ser Cys

15 TGG TAT CAA ACT TCT CTT ATA CTT CCC CCA GAA ACG ACC
Trp Tyr Gln Thr Ser Leu Ile Leu Pro Pro Glu Thr Thr

GGC CAG GTC TAC TTA TTA TCA TTC ATA AGT GCA TGT CCA
20 Gly Gln Val Tyr Leu Leu Ser Phe Ile Ser Ala Cys Pro

GAT TTT AAG CTT AGG CTG ATG AAA GAT ACT CAA ACT ATC
Asp Phe Lys Leu Arg Leu MET Lys Asp Thr Gln Thr Ile

25 VP3 VP1
TCA CAG ACT GTT GCA CTC ACT GAA GGC TTA GGT GAT GAA
Ser Gln Thr Val Ala Leu Thr Glu Gly Leu Gly Asp Glu

TTA GAA GAA GTC ATC GTT GAG AAA ACG AAA CAG ACG GTG
30 Leu Glu Glu Val Ile Val Glu Lys Thr Lys Gln Thr Val

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17112IA

GCC TCA ATC TCA TCT GGT CCA AAA CAC ACA CAA AAA GTC
Ala Ser Ile Ser Ser Gly Pro Lys His Thr Gln Lys Val

5 CCC ATA CTA ACT GCA AAC GAA ACA GGG GCC ACA ATG CCT
Pro Ile Leu Thr Ala Asn Glu Thr Gly Ala Thr MET Pro

GTT CTT CCA TCA GAC AGC ATA GAA ACC AGA ACT ACC TAC
Val Leu Pro Ser Asp Ser Ile Glu Thr Arg Thr Thr Tyr

10 ATG CAC TTT AAT GGT TCA GAA ACT GAT GTA GAA TGC TTT
MET His Phe Asn Gly Ser Glu Thr Asp Val Glu Cys Phe

TTG GGT CGT GCA GCT TGT GTG CAT GTA ACT GAA ATA CAA
Leu Gly Arg Ala Ala Cys Val His Val Thr Glu Ile Gln

15 AAC AAA GAT GCT ACT GGA ATA GAT AAT CAC AGA GAA GCA
Asn Lys Asp Ala Thr Gly Ile Asp Asn His Arg Glu Ala

AAA TTG TTC AAT GAT TGG AAA ATC AAC CTG TCC AGC CTT
20 Lys Leu Phe Asn Asp Trp Lys Ile Asn Leu Ser Ser Leu

GTC CAA CTT AGA AAG AAA CTG GAA CTC TTC ACT TAT GTT
Val Gln Leu Arg Lys Lys Leu Glu Leu Phe Thr Tyr Val

25 AGG TTT GAT TCT GAG TAT ACC ATA CTG GCC ACT GCA TCT
Arg Phe Asp Ser Glu Tyr Thr Ile Leu Ala Thr Ala Ser

CAA CCT GAT TCA GCA AAC TAT TCA AGC AAT TTG GTG GTC
Gln Pro Asp Ser Ala Asn Tyr Ser Ser Asn Leu Val Val

30

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CAA GCC ATG TAT GTT CCA CAT GGT GCC CCG AAA TCC AAA
Gln Ala MET Tyr Val Pro His Gly Ala Pro Lys Ser Lys

AGA GTG GGC GAT TAC ACA TGG CAA AGT GCT TCA AAC CCC
5 Arg Val Gly Asp Tyr Thr Trp Gln Ser Ala Ser Asn Pro

AGT GTA TTC TTC AAG GTG GGG GAT ACA TCA AGG TTT AGT
Ser Val Phe Phe Lys Val Gly Asp Thr Ser Arg Phe Ser

10 GTG CCT TAT GTA GGA TTG GCA TCA GCA TAT AAT TGT TTT
Val Pro Tyr Val Gly Leu Ala Ser Ala Tyr Asn Cys Phe

TAT GAT GGT TAC TCA CAT GAT GAT GCA GAA ACT CAG TAT
Tyr Asp Gly Tyr Ser His Asp Asp Ala Glu Thr Gln Tyr
15

GGC ATA ACT GTT CTA AAC CAT ATG GGT AGT ATG GCA TTC
Gly Ile Thr Val Leu Asn His MET Gly Ser MET Ala Phe

AGA ATA GTA AAT GAA CAT GAT GAA CAC AAA ACT CTT GTC
20 Arg Ile Val Asn Glu His Asp Glu His Lys Thr Leu Val

AAG ATC AGA GTT TAT CAC AGG GCA AAG CTC GTT GAA GCA
Lys Ile Arg Val Tyr His Arg Ala Lys Leu Val Glu Ala

25 TGG ATT CCA AGA GCA CCC AGA GCA CTA CCC TAC ACA TCA
Trp Ile Pro Arg Ala Pro Arg Ala Leu Pro Tyr Thr Ser

ATA GGG CGC ACA AAT TAT CCT AAG AAT ACA GAA CCA GTA
Ile Gly Arg Thr Asn Tyr Pro Lys Asn Thr Glu Pro Val
30

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VP1 3B

ATT AAG AAG AGG AAA GGT GAC ATT AAA TCC TAT GGT TTA
Ile Lys Lys Arg Lys Gly Asp Ile Lys Ser Tyr Gly Leu

5 GGA CCT AGG TAC GGT GGG ATT TAT ACA TCA AAT GTT AAA
Gly Pro Arg Tyr Gly Gly Ile Tyr Thr Ser Asn Val Lys

ATA ATG AAT TAC CAC TTG ATG ACA CCA GAA GAC CAC CAT
Ile MET Asn Tyr His Leu MET Thr Pro Glu Asp His His

10 AAT CTG ATA GCA CCC TAT CCA AAT AGA GAT TTA GCA ATA
Asn Leu Ile Ala Pro Tyr Pro Asn Arg Asp Leu Ala Ile

GTC TCA ACA GGA GGA CAT GGT GCA GAA ACA ATA CCA CAC
15 Val Ser Thr Gly Gly His Gly Ala Glu Thr Ile Pro His

TGT AAC CGT ACA TCA GGT GTT TAC TAT TCC ACA TAT TAC
Cys Asn Arg Thr Ser Gly Val Tyr Tyr Ser Thr Tyr Tyr

20 AGA AAG TAT TAC CCC ATA ATT TGC GAA AAG CCC ACC AAC ATC
Arg Lys Tyr Tyr Pro Ile Ile Cys Glu Lys Pro Thr Asn Ile

TGG ATT GAA GGA AGC CCT TAT TAC CCA AGT AGA TTT CAA
Trp Ile Glu Gly Ser Pro Tyr Tyr Pro Ser Arg Phe Gln

25 GCA GGA GTG ATG AAA GGG GTT GGG CCA GCA GAG CTA GGA
Ala Gly Val MET Lys Gly Val Gly Pro Ala Glu Leu Gly

GAC TGC GGT GGG ATT TTG AGA TGC ATA CAT GGT CCC ATT
30 Asp Cys Gly Gly Ile Leu Arg Cys Ile His Gly Pro Ile

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171121A

GGA TTG TTA ACA GCT GAA GGT AGT GGA TAT GTT TGT TTT
Gly Leu Leu Thr Ala Glu Gly Ser Gly Tyr Val Cys Phe

3B

5 GCT GAC ATA CGA CAG TTG GAG TGT ATC GCA GAG GAA CAG
Ala Asp Ile Arg Gln Leu Glu Cys Ile Ala Glu Glu Gln

5B

GGG CTG AGT GAT TAC ATC ACA GGT TTG GGT AGA GCT TTT
10 Gly Leu Ser Asp Tyr Ile Thr Gly Leu Gly Arg Ala Phe

GGT GTC GGG TTC ACT GAC CAA ATC TCA ACA AAA GTC ACA
Gly Val Gly Phe Thr Asp Gln Ile Ser Thr Lys Val Thr

15 GAA CTA CAA GAA GTG GCG AAA GAT TTC CTC ACC ACA AAA
Glu Leu Gln Glu Val Ala Lys Asp Phe Leu Thr Thr Lys

GTT TTG TCC AAA GTG GTC AAA ATG GTT TCA GCT TTA GTG
Val Leu Ser Lys Val Val Lys MET Val Ser Ala Leu Val

20

ATC ATT TGC AGA AAT CAT GAT GAC TTG GTC ACT GTT ACG
Ile Ile Cys Arg Asn His Asp Asp Leu Val Thr Val Thr

GCC ACT CTA GCA CTA CTT GGA TGT GAT GGA TCT CCT TGG
25 Ala Thr Leu Ala Leu Leu Gly Cys Asp Gly Ser Pro Trp

AGA TTT CTG AAG ATG TAC ATT TCC AAA CAC TTT CAG GTG
Arg Phe Leu Lys MET Tyr Ile Ser Lys His Phe Gln Val

30

5B X

CCT TAC ATT GAA AGA CAA GCA AAT GAT GGA TGG TTC AGA
Pro Tyr Ile Glu Arg Gln Ala Asn Asp Gly Trp Phe Arg

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17112IA

AAG TTT AAT GAT GCA TGT AAT GCT GCA AAG GGA TTG GAA
Lys Phe Asn Asp Ala Cys Asn Ala Ala Lys Gly Leu Glu

5 TGG ATT GCT AAT AAG ATT TCC AAA CTG ATT GAA TGG ATA
Trp Ile Ala Asn Lys Ile Ser Lys Leu Ile Glu Trp Ile

AAA AAC AAA GTA CTT CCC CAA GCC AAA GAA AAA CTA GAA
Lys Asn Lys Val Leu Pro Gln Ala Lys Glu Lys Leu Glu

10 TTT TGT AGT AAA CTC AAA CAA CTT GAT ATA CTA GAG AGA
Phe Cys Ser Lys Leu Lys Gln Leu Asp Ile Leu Glu Arg

CAA ATA ACC ACC ATG CAT ATC TCG AAT CCA ACA CAG GAA
Gln Ile Thr Thr MET His Ile Ser Asn Pro Thr Gln Glu

15 AAA CGA GAG CAG TTG TTC AAT AAC GTA TTG TGG TTG GAA
Lys Arg Glu Gln Leu Phe Asn Asn Val Leu Trp Leu Glu

CAA ATG TCG CAA AAG TTT GCC CCA TTT TAT GCC GTT GAA
20 Gln MET Ser Gln Lys Phe Ala Pro Phe Tyr Ala Val Glu

TCA AAA AGA ATC AGG GAA CTC AAG AAC AAA ATG GTA AAT
Ser Lys Arg Ile Arg Glu Leu Lys Asn Lys MET Val Asn

25 TAT ATG CAA TTT AAA AGT AAA CAA AGA ACT GAA CCA GTG
Tyr MET Gln Phe Lys Ser Lys Gln Arg Thr Glu Pro Val

TGT GTA TTA ATC CAT GGT ACA CCC GGT TCT GGT AAA TCA
Cys Val Leu Ile His Gly Thr Pro Gly Ser Gly Lys Ser

30 TTA ACA ACA TCC ATT GTG GGA CGT GCA ATT GCA GAA CAC
Leu Thr Thr Ser Ile Val Gly Arg Ala Ile Ala Glu His

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17112IA

TTC AAT TCA GCA GTA TAT TCA CTT CCA CCA GAT CCC AAG
Phe Asn Ser Ala Val Tyr Ser Leu Pro Pro Asp Pro Lys

5 CAC TTT GAT GGT TAT CAG CAA CAG GAA GTT GTG ATT ATG
His Phe Asp Gly Tyr Gln Gln Gln Glu Val Val Ile MET

GAT GAT CTG AAC CAA AAT CCA GAT GGA CAG GAT ATA AGC
Asp Asp Leu Asn Gln Asn Pro Asp Gly Gln Asp Ile Ser

10 ATG TTT TGT CAA ATG GTT TCT TCA GTG GAT TTC TTG CCT
MET Phe Cys Gln MET Val Ser Ser Val Asp Phe Leu Pro

CCA ATG GCT AGT TTA GAT AAC AAG GGC ATG TTA TTC ACC
Pro MET Ala Ser Leu Asp Asn Lys Gly MET Leu Phe Thr

15 AGT AAT TTT GTT CTA GCC TCC ACA AAT TCT AAC ACA CTA
Ser Asn Phe Val Leu Ala Ser Thr Asn Ser Asn Thr Leu

AGC CCC CCA ACA ATC TTG AAT CCT GAA GCT TTA GTC AGG
20 Ser Pro Pro Thr Ile Leu Asn Pro Glu Ala Leu Val Arg

AGA TTT GGT TTT GAC CTA GAT ATA TGT TTG CAT ACT ACC
Arg Phe Gly Phe Asp Leu Asp Ile Cys Leu His Thr Thr

25 TAC ACA AAG AAT GGA AAA CTC AAT GCA GGC ATG TCA ACC
Tyr Thr Lys Asn Gly Lys Leu Asn Ala Gly MET Ser Thr

AAG ACA TGC AAA GAT TGC CAT CAA CCA TCT AAT TTC AAG
Lys Thr Cys Lys Asp Cys His Gln Pro Ser Asn Phe Lys

30

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17112IA

AAA TGT TGC CCC CTA GTC TGT GGA AAA GCT ATT AGC TTG
Lys Cys Cys Pro Leu Val Cys Gly Lys Ala Ile Ser Leu

5 GTA GAC AGA ACT ACC AAC GTT AGG TAT AGT GTG GAT CAA
Val Asp Arg Thr Thr Asn Val Arg Tyr Ser Val Asp Gln

CTG GTC ACG GCT ATT ATA AGT GAT TTC AAG AGC AAA ATG
Leu Val Thr Ala Ile Ile Ser Asp Phe Lys Ser Lys MET

10 X 1B
CAA ATT ACA GAT TCC CTA GAA ACA CTG TTT CAA GGA CCA
Gln Ile Thr Asp Ser Leu Glu Thr Leu Phe Gln Gly Pro

15 GTG TAT AAA GAT TTA GAG ATT GAT GTT TGC AAC ACA CCA
Val Tyr Lys Asp Leu Glu Ile Asp Val Cys Asn Thr Pro

CCT TCA GAA TGT ATC AAC GAT TTA CTG AAA TCT GTA GAT
Pro Ser Glu Cys Ile Asn Asp Leu Leu Lys Ser Val Asp

20 TCA GAA GAG ATT AGG GAA TAT TGT AAG AAG AAG AAA TGG
Ser Glu Glu Ile Arg Glu Tyr Cys Lys Lys Lys Lys Trp

ATT ATA CCT GAA ATT CCT ACC AAC ATA GAA AGG GCT ATG
Ile Ile Pro Glu Ile Pro Thr Asn Ile Glu Arg Ala MET

25 AAT CAA GCC AGC ATG ATT ATT AAT ACT ATT CTG ATG TTT
Asn Gln Ala Ser Met Ile Ile Asn Thr Ile Leu MET Phe

30

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17112IA

GTC AGT ACA TTA GGT ATT GTT TAT GTC ATT TAT AAA TTG
Val Ser Thr Leu Gly Ile Val Tyr Val Ile Tyr Lys Leu

1B VPg

5 TTT GCT CAA ACT CAA GGA CCA TAT TCT GGT AAC CCG CCT
Phe Ala Gln Thr Gln Gly Pro Tyr Ser Gly Asn Pro Pro

CAC AAT AAA CTA AAA GCC CCA ACT TTA CGC CCA GTT GTT
His Asn Lys Leu Lys Ala Pro Thr Leu Arg Pro Val Val

10

VPg Protease

GTG CAA GGA CCA AAC ACA GAA TTT GCA CTA TCC CTG TTA
Val Gln Gly Pro Asn Thr Glu Phe Ala Leu Ser Leu Leu

15 AGG AAA AAC ATA ATG ACT ATA ACA ACC TCA AAG GGA GAG
Arg Lys Asn Ile MET Thr Ile Thr Thr Ser Lys Gly Glu

TTC ACA GGG TTA GGC ATA CAT GAT CGT GTC TGT GTG ATA
Phe Thr Gly Leu Gly Ile His Asp Arg Val Cys Val Ile

20

CCC ACA CAC GCA CAG CCT GGT GAT GAT GTA CTA GTG AAT
Pro Thr His Ala Gln Pro Gly Asp Asp Val Leu Val Asn

GGT CAG AAA ATT AGA GTT AAG GAT AAG TAC AAA TTA GTA
25 Gly Gln Lys Ile Arg Val Lys Asp Lys Tyr Lys Leu Val

GAT CCA GAG AAC ATT AAT CTA GAG CTT ACA GTG TTG ACT
Asp Pro Glu Asn Ile Asn Leu Glu Leu Thr Val Leu Thr

30 TTA GAT AGA AAT GAA AAA TTC AGA GAT ATC AGG GGA TTT
Leu Asp Arg Asn Glu Lys Phe Arg Asp Ile Arg Gly Phe

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ATA TCA GAA GAT CTA GAA GGT GTG GAT GCC ACT TTG GTA
Ile Ser Glu Asp Leu Glu Gly Val Asp Ala Thr Leu Val

5 GTA CAT TCA AAT AAC TTT ACC AAC ACT ATC TTA GAA GTT
Val His Ser Asn Asn Phe Thr Asn Thr Ile Leu Glu Val

GGC CCT GTA ACA ATG GCA GGA CTT ATT AAT TTG AGT AGC
Gly Pro Val Thr MET Ala Gly Leu Ile Asn Leu Ser Ser

10 ACC CCC ACT AAC AGA ATG ATT CGT TAT GAT TAT GCA ACA
Thr Pro Thr Asn Arg MET Ile Arg Tyr Asp Tyr Ala Thr

AAA ACT GGG CAG TGT GGA GGT GTG CTG TGT GCT ACT GGT
Lys Thr Gly Gln Cys Gly Gly Val Leu Cys Ala Thr Gly
15

AAG ATC TTT GGT ATT CAT GTT GGC GGT AAT GGA AGA CAA
Lys Ile Phe Gly Ile His Val Gly Gly Asn Gly Arg Gln

20 GGA TTT TCA GCT CAA CTT AAA AAA CAA TAT TTT GTA GAG
Gly Phe Ser Ala Gln Leu Lys Lys Gln Tyr Phe Val Glu

Protease Replicase

AAA CAA GGC CAA GTA ATA GCT AGA CAT AAG GTT AGG GAG
Lys Gln Gly Gln Val Ile Ala Arg His Lys Val Arg Glu
25

TTT AAC ATA AAT CCA GTC AAC ACG GCA ACT AAG TCA AAA
Phe Asn Ile Asn Pro Val Asn Thr Ala Thr Lys Ser Lys

30 TTA CAT CCC AGT GTA TTT TAT GAT GTT TTT CCA GGT GAC
Leu His Pro Ser Val Phe Tyr Asp Val Phe Pro Gly Asp

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AAG GAA CCT GCT GTA TTG AGT GAC AAT GAT CCC AGA CTG
Lys Glu Pro Ala Val Leu Ser Asp Asn Asp Pro Arg Leu

5 GAA GTT AAA TTG ACT GAA TCA TTA TTC TCT AAG TAC AAG
Glu Val Lys Leu Thr Glu Ser Leu Phe Ser Lys Tyr Lys

GGG AAT GTA AAT ACG GAA CCC ACT GAA AAT ATG CTT GTG
Gly Asn Val Asn Thr Glu Pro Thr Glu Asn MET Leu Val

10 GCT GTA GAC CAT TAT GCA GGG CAA CTA TTA TCA CTA GAT
Ala Val Asp His Tyr Ala Gly Gln Leu Leu Ser Leu Asp

ATC CCC ACT TCT GAA CTT ACA CTA AAA GAA GCA TTA TAT
Ile Pro Thr Ser Glu Leu Thr Leu Lys Glu Ala Leu Tyr

15 GGA GTA GAT GGA CTA GAA CCT ATA GAT ATT ACA ACC AGT
Gly Val Asp Gly Leu Glu Pro Ile Asp Ile Thr Thr Ser

GCA GGA TTT CCC TAT GTG AGT CTT GGG ATC AAA AAG AGA
20 Ala Gly Phe Pro Tyr Val Ser Leu Gly Ile Lys Lys Arg

GAC ATT CTG AAT AAA GAG ACC CAG GAC ACA GAA AAG ATG
Asp Ile Leu Asn Lys Glu Thr Gln Asp Thr Glu Lys MET

25 AAG TTT TAT CTA GAC AAG TAT GGC ATT GAC TTG CCT CTA
Lys Phe Tyr Leu Asp Lys Tyr Gly Ile Asp Leu Pro Leu

GTT ACA TAT ATT AAG GAT GAA TTA AGA AGT GTT GAC AAA
Val Thr Tyr Ile Lys Asp Glu Leu Arg Ser Val Asp Lys

30

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GTC CGA TTA GGG AAA AGT AGA TTA ATT GAA GCC TCC AGT
Val Arg Leu Gly Lys Ser Arg Leu Ile Glu Ala Ser Ser

5 TTG AAT GAT TCT GTT AAC ATG AGA ATG AAA CTA GGC AAC
Leu Asn Asp Ser Val Asn MET Arg MET Lys Leu Gly Asn

CTT TAC AAA GCA TTC CAT CAA AAT CCC GGT GTT CTG ACT
Leu Tyr Lys Ala Phe His Gln Asn Pro Gly Val Leu Thr

10 GGA TCA GCA GTG GGT TGT GAT CCT GAT GTG TTT TGG TCT
Gly Ser Ala Val Gly Cys Asp Pro Asp Val Phe Trp Ser

GTC ATC CCT TGC TTA ATG GAT GGG CAC CTG ATG GCA TTT
Val Ile Pro Cys Leu MET Asp Gly His Leu MET Ala Phe

15 GAT TAC TCT AAT TTT GAT GCC TCT TTG TCA CCA GTT TGG
Asp Tyr Ser Asn Phe Asp Ala Ser Leu Ser Pro Val Trp

TTT GTC TGT CTA GAG AAG GTT TTG ACC AAG TTA GGC TTT
20 Phe Val Cys Leu Glu Lys Val Leu Thr Lys Leu Gly Phe

GCA GGC TCT TCA TTA ATT CAA TCA ATT TGT AAT ACC CAT
Ala Gly Ser Ser Leu Ile Gln Ser Ile Cys Asn Thr His

25 CAT ATC TTT AGG GAT GAA ATA TAT GTG GTT GAA GGT GGC
His Ile Phe Arg Asp Glu Ile Tyr Val Val Glu Gly Gly

ATG CCC TCA GGG TGT TCA GGA ACC AGC ATA TTC AAT TCC
MET Pro Ser Gly Cys Ser Gly Thr Ser Ile Phe Asn Ser

30 ATG ATC AAC AAC ATA ATC ATT AGG ACT TTG ATA TTA GAT
MET Ile Asn Asn Ile Ile Ile Arg Thr Leu Ile Leu Asp

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GCA TAT AAA GGA ATA GAT TTA GAC AAA CTT AAA ATC TTA
Ala Tyr Lys Gly Ile Asp Leu Asp Lys Leu Lys Ile Leu

5 GCT TAC GGT GAT GAT TTG ATT GTT TCT TAT CCT TAT GAA
Ala Tyr Gly Asp Asp Leu Ile Val Ser Tyr Pro Tyr Glu
CTG GAT CCA CAA GTG TTG GCA ACT CTT GGT AAA AAT TAT
Leu Asp Pro Gln Val Leu Ala Thr Leu Gly Lys Asn Tyr

10 GGA CTA ACC ATC ACA CCC CCA GAC AAA TCT GAA ACT TTT
Gly Leu Thr Ile Thr Pro Pro Asp Lys Ser Glu Thr Phe

ACA AAA ATG ACA TGG GAA AAC TTG ACA TTT TTA AAG AGA
Thr Lys MET Thr Trp Glu Asn Leu Thr Phe Leu Lys Arg

15 TAC TTC AAG CCT GAT CAA CAA TTT CCC TTT TTG GTT CAC
Tyr Phe Lys Pro Asp Gln Gln Phe Pro Phe Leu Val His

CCA GTT ATG CCC ATG AAA GAT ATA CAT GAG TCA ATC AGA
Pro Val MET Pro MET Lys Asp Ile His Glu Ser Ile Arg

20 TGG ACA AAG GAT CCT AAA AAC ACA CAG GAT CAC GTC CGA
Trp Thr Lys Asp Pro Lys Asn Thr Gln Asp His Val Arg

TCA TTA TGC ATG TTA GCA TGG CAC TCA GGA GAA AAA GAG
25 Ser Leu Cys MET Leu Ala Trp His Ser Gly Glu Lys Glu

TAC AAT GAA TTC ATT CAG AAG ATC AGA ACT ACT GAC ATT
Tyr Asn Glu Phe Ile Gln Lys Ile Arg Thr Thr Asp Ile

30 GGA AAA TGT CTA ATT CTC CCA GAA TAC AGC GTA CTT AGG
Gly Lys Cys Leu Ile Leu Pro Glu Tyr Ser Val Leu Arg

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Replicase

AGG CGC TGG TTG GAC CTC TTT

Arg Arg Trp Leu Asp Leu Phe

5 TAGGTTAACAATATAGACACTTAATTTGAGTAGAAGTAGGAGT

TTATAAAAAA AAAAAAAA

10 Nucleotide and corresponding amino acid
sequences for the various structural and non-
structural HRV14 proteins are indicated. Nomen-
clature employed is based upon that used for the
polioviruses. Neutralizing epitopes to HRV14 have
15 been mapped to each of the VP1, VP2 and VP3 virus
structural proteins. The virus protease is required
for proper proteolytic cleavage of the polyprotein;
inhibition of its activity (e.g. by Zn^{++}) results
in blockage of virus replication. Virus coded
20 replicase and VP-g is required for synthesis of both
plus and minus strands of genomic RNA; inhibition of
its activity prevents virus replication.

EXAMPLE 11Immunization of Mice with Purified Rhinovirus Type 14

25 The rear footpads of adult BALB/C mice were
injected with purified whole HRV-14 as obtained
according to Example 1. Each mouse footpad received
5 µg of purified HRV-14 dissolved in a volume of
0.1 ml of complete Freund's adjuvant. Mice received
30 a second inoculation (5 µg/0.05 ml/footpad) at 30
days. Serum was collected from one eye of each mouse
14 days after viral injection and tested for antibody

to HRV-14 by neutralization assay (Example 14).

Mice that were positive for anti-HRV-14 antibodies by neutralization assays (as described in Example 14) were primed 3 days prior to the cell fusion by injection of 5 µg of purified HRV-14 in the tail vein (in aqueous solution, 0.1 ml/mouse). Sera collected at death from the two animals used for the cell fusion were assayed for antibody to HRV-14 by neutralization and found to be positive.

10

EXAMPLE 12

Immunization of Mice with HeLa R19 Cells

Adult BALB/C mice were injected intraperitoneally with a solution containing HeLa R19 cells (Example 1) which were recovered from tissue culture monolayers by treatment with phosphate buffered saline (PBS) containing 50 mM EDTA for 20 mins. at 37°C. Each mouse received 3×10^6 cells suspended in a volume of 0.5 ml of complete Freund's adjuvant. After 38 days, mice were reinoculated intraperitoneally with 1×10^7 HeLa R19 cells in a volume of 0.5 ml of incomplete Freund's adjuvant.

Three mice that were positive for antibodies which protected cells against HRV-14 infection (See Example 14) were primed on day 116, 4 days prior to the cell fusion by injection of two OD₂₆₀ units/mouse of crude HeLa cell membranes (See Example 13) in the tail vein (in aqueous solution, 0.05 ml mouse). Sera collected at the death of the three animals used showed the presence of antibody capable of protecting HeLa cells from HRV-14 infection.

In another experiment three adult BALB/C mice were immunized using the following schedule: 2 intraperitoneal inoculations using cell membrane preparations on day 1 and day 42 followed by a final
5 intradermal inoculation of membranes plus incomplete Freund's adjuvant on day 214. Three days later the cell fusion was carried out as described in Example 14.

EXAMPLE 13

Preparation of HeLa Cell Membranes

HeLa R19 cell monolayers were treated with PBS containing 50 mM EDTA for 20 minutes at 37°C. Cells were collected by low speed centrifugation and washed with PBS 3 times. Cells were pelleted and
15 resuspended in 10 mM phosphate buffer (8×10^7 cells/ml) and placed on ice for 25 mins. Cells were then disrupted by 20 stokes in a Dounce homogenizer and nuclei removed by centrifugation (1000 x g for 5 minutes at 4°C). The supernatant liquid was removed
20 and membranes pelleted at 143,000 x g for 1 hour at 4°C in a Beckman TI60 rotor. The supernatant liquid was discarded and the crude membrane pellet dissolved in PBS at 42 OD₂₈₀ units/ml and stored at -70°C.

EXAMPLE 14

Production of Hybridomas by Fusion of Immune Mouse Spleen Cells with Mouse Myeloma Cells

Mouse myeloma cells (SP 2/0) were grown from frozen seed stock in HT media. Each 100 ml of HT
30 media contained 66 ml of Dulbecco's Modified Eagle's Media; 20 ml fetal calf serum (FCS); 10 ml of NCTC 109 with Eagle's balanced salts; 2 ml of 200 mM

L-glutamine; 1 ml of a solution containing 408 mg hypoxanthine and 114 mg thymidine in 300 ml of distilled H₂O; and 1 ml of OPI stock (1.5 g cis-oxalacetic acid, 0.5 g pyruvic acid, 2000 units bovine insulin in 100 ml H₂O); and 0.2 ml of a penicillin (10,000 units/ml) and streptomycin (10,000 µg/ml) mixture. The mouse spleens from animals immunized with HRV-14 (Example 12) were removed after cervical dislocation and placed in serumless HT media at room temperature. The spleens were placed in a plastic petri dish and the cells were teased from the spleen with a plastic scraper. The plates were washed with 5 ml of serumless HT media and the cells were pooled into a 15 ml plastic conical tube; large particles were allowed to settle for one minute. The supernatant was then transferred to a 15 ml plastic round bottom tube and the cells were pelleted by a 10 minute centrifugation, 350 x g at room temperature. The supernatant liquid was discarded and the cells were resuspended in serumless HT media (10 ml/2 spleens) and the total viable cells were determined by trypan blue exclusion.

The number of viable SP 2/0 myeloma cells was also determined and the cells were combined using one log more spleen cells than myeloma cells (i.e. into a 50 cc screw cap plastic tube were placed 2×10^8 spleen cells and 2×10^7 SP 2/0 cells). The cells were pelleted by centrifugation for 10 minutes at 350 x g and then the cell pellet was resuspended in 10 ml of serumless HT media. This pelleting and resuspension-washing procedure was repeated two more times, and after the final

resuspension in 10 ml of serumless HT media the cells were transferred to a 15 ml round bottom tube. The cells were pelleted at 350 x g for 10 minutes.

Polyethylene glycol (PEG; molecular weight average = 1000d) was liquified at 45°C and combined with serumless HT media to a concentration of 35% PEG (vol/vol). The PEG/HT media mixture was sterilized by passing it through a 0.22 micron membrane filter fitted on the end of 3 ml syringe; the PEG was then maintained at 37°C. Dimethylsulfoxide (DMSO) was added to the PEG/media mixture to a final concentration of 5%. The PEG/DMSO/HT media mixture was added dropwise to the spleen-myeloma cell pellett, using 0.8 ml for 2×10^8 cells while gently resuspending the cells by tapping the side of the culture tube and swirling the cells. The cell pellets were centrifuged at 250 x g at room temperature so that the total contact time of the cells with PEG was 6 minutes. The PEG supernatant was then aspirated off and the cells were resuspended in 10 ml of HT media. The cells were pelleted (350 x g for 10 minutes) and gently resuspended in HT media to a final concentration of 3.5×10^5 myeloma cells/ml. The cells were then plated in 96 well microtiter plates using 0.1 ml/well with a pipet and incubated at 37°C in a water-jacketed CO₂ incubator with 5% CO₂ and 96% humidity.

After 24 hours, 0.1 ml of HAT media (HT media plus aminopterin at 0.352 mg/liter) was added to each well. The wells were refed with 0.1 ml of fresh HAT media every 4 to 5 days.

Some cells from culture wells which were at a confluency of 15% or greater (usually 40 to 80%) were tested for production of antibodies to HRV-14 or by a neutralization assay described in Example 15.

5 In other experiments some cells from growth-positive wells which were at a confluency of 15% or greater were tested for production of antibodies which block attachment of HRV-14 to cellular receptors in a cell protection assay
10 described in Example 15.

From the first set of three mice in Example 12, 837 growth-positive wells were obtained, and from the second set of three mice in Example 12, 693 growth-positive wells were obtained. Screening of
15 culture fluids from the total 1530 wells resulted in the identification of two hybridoma cultures, one from each set of three mice, producing antibodies capable of blocking the attachment of the major group of human rhinoviruses to susceptible cells.

20 Those cells having high activity by the above assays were subcloned by two cycles of limiting dilutions. Eight subclones (A-H) were selected which

25

30

had high HRV-14 neutralization activities and one subclone (I) which was selected in a cell protection assay. The purified monoclonal antibodies from these subclones were characterized as to immunoglobulin subtype and molecular weight by standard procedure. The following results were obtained:

	MONOCLONAL ANTIBODY	SUBTYPE	MOLECULAR WEIGHT (Daltons)	
			Heavy Chain	Light Chain
10	A	IgG-2A	49,000	27,000
	B	IgG-2A	49,000	27,000
	C	IgG-2A	49,000	27,000
	D	IgG-2A	49,000	27,000
	E	IgG-1	49,000	27,000
15	F	IgG-2A	49,000	27,000
	G	IgG-2A	49,000	27,000
	H	IgG-2A	49,000	27,000
	I	IgG-1	49,000	27,000

20

EXAMPLE 15Neutralization/Cell Protection Assays

Neutralization assays were performed as follows: HRV-14 (10^6 PFU) was incubated with 40 μ l of serum or hydridoma tissue culture fluid in a total volume of 0.2 ml GM for 1 hour at 25°C. The solution was then transferred to a HeLa R19 cell monolayer (2.5×10^5 cells) in a 24-well cluster plate and incubated 16-24 hours at 34°C in a CO₂ incubator. Wells were scored as positive if no evidence of cytopathic effect was evident compared to controls.

30

Protection assays utilized 48-well cluster plates of HeLa R-19 cells (1.25×10^5 cells/well).

After removal of media, monolayers were incubated with 0.1 ml serum or hybridoma tissue culture for 1 hour at 34°C. After incubation, 2×10^5 PFUs of HRV-14 in 50 μ l of media were added to the treated
5 monolayer and the cells incubated 16-24 hours at 34°C in a CO₂ incubator. Wells were scored as positive if no evidence of cytopathic effect was evident compared to controls. Further assays were done to determine if this HeLa receptor antibody would
10 protect cells against infection with other serotypes of HRVs. Positive protection results were obtained with HRV types 3, 5, 9, 11, 12, 14, 15, 17, 22, 32, 36, 39, 41, 51, 58, 59, 60, 66, 67, 89, and a clinical isolate of unknown serotype, and coxsackie
15 virus types A13, A18 and A21. All of these serotypes are known to share the same cellular receptor. Four serotypes of rhinovirus which share a different receptor (Types 1A, 2, 44, and 49) were not blocked by the antibody. This monoclonal antibody also
20 failed to protect cells against Sabin Type 1 polio virus and coxsackie viruses Types B2 and B3.

EXAMPLE 16

Purification of Monoclonal antibodies from 25 Tissue Culture Media and Ascites Fluid

The monoclonal antibodies used in experiments were purified from tissue culture fluid taken from growing cultures of the hybridoma cells producing individual antibody. The following
30 procedure was employed and is essentially that described by Emini et al. (J. Virol. 43:997-1005, 1982). Tissue culture fluid, 1 liter, from a

particular hybridoma cell line was filtered through a Whatmann 1MM filter paper, a glass wool containing column, and finally a column containing agarose (Sephadex 6B). The tissue culture fluid was then
5 adjusted to a pH of 8.0 by the addition of 100 mM tris-HCl, pH 8.0, and then passed over a protein A-Sepharose column (4 to 6 ml bed volume per 2 liters of fluid). The column was washed with 10 bed volumes of 10 mM Tris-HCl (pH 8.0). Glycine (100 mM) pH 3.0
10 was then added to elute the bound immunoglobulins from the column; fractions were collected with 100 mM Tris-HCl (pH 8.0) in the bottom of each tube such that the pH was changed from the acid pH of glycine to the higher pH to stabilize the antibody. Peak
15 protein fractions from the column were pooled, dialyzed against distilled H₂O and used as whole antibody or were treated further to obtain Fab fragments.

To obtain Fab fragments of the individual
20 monoclonal antibodies, 10 to 15 mg of the antibody purified using protein A columns, as described above, were lyophilized and then dissolved in 100 mM sodium phosphate, 10 mM cysteine, pH 7.2 (2 ml/20 mg antibody). Papain, 10-15 units per mg, was added to
25 a ratio of 1:100, enzyme to antibody by weight. This reaction was incubated overnight for 16 hours at 37°C and it was terminated with the addition of iodoacetamide to a final concentration of 30 mM. The digested antibody was then chromatographed over a
30 cross-linked dextran (Sephadex G-75) column (2.5 X 20 cm, width x height) at 4°C and the fractions were monitored for absorbance at 280 nm as a measure of

the protein present. The second peak off the column, which contained the majority of the Fab fragments, was pooled and then passed through a protein A-sepharose column to eliminate any contaminating Fc fragments or undigested antibody which bind to the protein A column. The peak fractions of Fab fragments from the protein A column were pooled, dialyzed against distilled H₂O, and then lyophilized in 200 µg aliquots and stored at -80°C.

10

EXAMPLE 17

Crosslinking Monoclonal Antibodies to HRV

Neutralizing monoclonal antibodies directed towards HRV14 were cross-linked to the virus by the use of the heterobifunctional crosslinker toluene-2,4-diisocyanate (TDI), with the following minor adaptations of the procedure described in Emini *et al.* (J. Virol. 43:997-1005, 1982). Sodium phosphate buffer (10 mM, pH 7.2) was added to dried Fab fragments of a monoclonal antibody, usually to a final concentration of 0.5 µg/µl. TDI was then added, 1.1 µl/200 µl of antibody. The antibody-Fab fragment was then incubated with the TDI at either 4°C or at room temperature. The reaction was terminated by taking aliquots at various times, usually less than 10 minutes, and incubating these aliquots at 0°C for 10 minutes to solidify the TDI. The TDI was removed by pelleting the solidified TDI at 2,000 x g at 0°C and then repeating the 0°C incubation and pelleting steps. The final supernatant liquid then was taken and HRV14 was added to the antibody. HRV was added either as S³⁵ labelled HRV, with 3000 to 7000 cpm.

The virus was incubated with the antibody for 30 minutes at room temperature.

Saturated tribasic phosphate buffer was added to raise the pH to 9.6 for 10 minutes at room temperature. Saturated monobasic phosphate was then added to lower the pH to 7.0. The samples then were dialyzed against dH_2O overnight with 3 changes of the water. Each sample was lyophilized to dryness and run on a polyacrylamide gel (Laemmli, Nature 227: 680-685, 1970). Following the electrophoresis, the gel was dried and then exposed to X-ray film for varying lengths of time. To determine the degree of crosslinking to individual proteins, a densitometer tracing of the X-ray film was performed which also gave a quantitative measure of the various viral peaks. The neutralizing monoclonal antibodies were found to crosslink to VP1, VP2 or VP3.

20

25

30

WHAT IS CLAIMED IS:

1. DNA representative of genomic RNA of HRV
Type 14 having the nucleotide and corresponding amino
acid sequence:
- 5 TTAAACAGCG GATGGGTATC CCACCATTCG ACCCATTTGGG TGTAGTACTC
TGGTACTATG TACCTTTGTA CGCCTGTTTC TCCCCAACCA CCCTTCCTTA
AAATTCCCAC CCATGAAACG TTAGAAGCTT GACATTAAAG TACAATAGGT
GGCGCCATAT CCAATGGTGT CTATGTACAA GCACTTCTGT TTCCCAGGAG
10 CGAGGTAAAG GCTGTACCCA CTGCCAAAAG CCTTTAACCG TATCCGCCAA
CCAAC TACGT AACAGTTAGT ACCATCTTGT TCTTGACTGG ACGTTCGATC
AGGTGGATTT TCCCTCCACT AGTTTGGTCTG ATGAGGCTAG GAATTCCTCA
CGGGTGACCG TGTCTTAGCC TCGGTGGCGG CCAGACCCAG CTTATGCTGG
GACGCCCTTT TAAGGACATG GTGTGAAGAC TCGCATGTGC TTGGTTGTGA
15 GTCCTCCGGC CCCTGAATGC GGCTAACCTT AACCCTAGAG CCTTATGCCA
CGATCCAGTG GTTGTAAGGT CGTAATGAGC AATTCCGGGA CGGGACCGAC
TACTTTGGGT GTCCGTGTTT CTCATTTTTC TTCATATTGT CTTATGGTCA
CAGCATATAT ATACATATAC TGTGATC
- 20 VP4
ATG GGC GCT CAG GTT TCT ACA CAG AAA AGT GGA TCT CAC
MET Gly Ala Gln Val Ser Thr Gln Lys Ser Gly Ser His
- GAA AAT CAA AAC ATT TTG ACC AAT GGA TCA AAT CAG ACT
25 Glu Asn Gln Asn Ile Leu Thr Asn Gly Ser Asn Gln Thr
- TTC ACA GTT ATA AAT TAC TAT AAG GAT GCA GCA AGT ACA
Phe Thr Val Ile Asn Tyr Tyr Lys Asp Ala Ala Ser Thr
- 30 TCA TCA GCT GGT CAA TCA CTG TCA ATG GAC CCA TCT AAG
Ser Ser Ala Gly Gln Ser Leu Ser MET Asp Pro Ser Lys

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TTT ACA GAA CCA GTT AAA GAT CTC ATG CTT AAG GGT GCA
Phe Thr Glu Pro Val Lys Asp Leu MET Leu Lys Gly Ala

VP4 VP2

5 CCA GCA TTG ATT TCA CCC AAT GTT GAG GCC TGT GGT TAT
Pro Ala Leu Asn Ser Pro Asn Val Glu Ala Cys Gly Tyr

AGT GAT AGA GTA CAA CAA ATC ACA CTC GGG AAT TCA ACA
Ser Asp Arg Val Gln Gln Ile Thr Leu Gly Asn Ser Thr

10

ATA ACA ACA CAA GAA GCA GCC AAC GCT GTT GTG TGT TAT
Ile Thr Thr Gln Glu Ala Ala Asn Ala Val Val Cys Tyr

15 GCT GAA TGG CCA GAG TAC CTT CCA GAT GTG GAC GCT AGT
Ala Glu Trp Pro Glu Tyr Leu Pro Asp Val Asp Ala Ser

GAT GTC AAT AAA ACT TCA AAA CCA GAC ACT TCT GTC TGT
Asp Val Asn Lys Thr Ser Lys Pro Asp Thr Ser Val Cys

20 AGG TTT TAC ACA TTG GAT AGT AAG ACA TGG ACA ACA GGT
Arg Phe Tyr Thr Leu Asp Ser Lys Thr Trp Thr Thr Gly

TCT AAA GGC TGG TGC TGG AAA TTA CCA GAT GCA CTC AAG
Ser Lys Gly Trp Cys Trp Lys Leu Pro Asp Ala Leu Lys

25

GAT ATG GGT GTG TTC GGG CAA AAC ATG TTT TTC TAC TCA
Asp MET Gly Val Phe Gly Gln Asn MET Phe Phe Tyr Ser

30 CTA GGA AGA TCA GGT TAC ACA GTA CAC GTT CAG TGC AAT
Leu Gly Arg Ser Gly Tyr Thr Val His Val Gln Cys Asn

GCC ACA AAA TTC CAT AGC GGT TGT CTA CTT GTA GTT GTA
Ala Thr Lys Phe His Ser Gly Cys Leu Leu Val Val Val

5 ATA CCA GAA CAC CAA CTG GCT TCA CAT GAG GGT GGC AAT
Ile Pro Glu His Gln Leu Ala Ser His Glu Gly Gly Asn

GTT TCA GTT AAA TAC ACA TTC ACG CAT CCA GGT GAA CGT
Val Ser Val Lys Tyr Thr Phe Thr His Pro Gly Glu Arg

10 GGT ATA GAT TTA TCA TCT GCA AAT GAA GTG GGA GGG CCT
Gly Ile Asp Leu Ser Ser Ala Asn Glu Val Gly Gly Pro

GTC AAG GAT GTC ATA TAC AAT ATG AAT GGT ACT TTA TTA
15 Val Lys Asp Val Ile Tyr Asn MET Asn Gly Thr Leu Leu

GGA AAT CTG CTC ATT TTC CCT CAC CAG TTC ATT AAT CTA
Gly Asn Leu Leu Ile Phe Pro His Gln Phe Ile Asn Leu

20 AGA ACC AAT AAT ACA GCC ACA ATA GTG ATA CCA TAC ATA
Arg Thr Asn Asn Thr Ala Thr Ile Val Ile Pro Tyr Ile

AAC TCA GTA CCC ATT GAT TCA ATG ACA CGT CAC AAC AAT
Asn Ser Val Pro Ile Asp Ser MET Thr Arg His Asn Asn

25 GTC TCA CTG ATG GTC ATC CCT ATT GCC CCT CTT ACA GTA
Val Ser Leu MET Val Ile Pro Ile Ala Pro Leu Thr Val

CCA ACT GGA GCA ACT CCC TCA CTC CCT ATA ACA GTC ACA
30 Pro Thr Gly Ala Thr Pro Ser Leu Pro Ile Thr Val Thr

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ATA GCA CCT ATG TGC ACT GAG TTC TCT GGG ATA AGG TCC
Ile Ala Pro MET Cys Thr Glu Phe Ser Gly Ile Arg Ser

VP2 VP3

5 AAG TCA ATT GTG CCA CAA GGT TTG CCA ACT ACA ACT TTG
Lys Ser Ile Val Pro Gln Gly Leu Pro Thr Thr Thr Leu

CCG GGG TCA GGA CAA TTC TTG ACC ACA GAT GAC AGG CAA
Pro Gly Ser Gly Gln Phe Leu Thr Thr Asp Asp Arg Gln

10

TCC CCC AGT GCA CTG CCA AAT TAT GAG CCA ACT CCA AGA
Ser Pro Ser Ala Leu Pro Asn Tyr Glu Pro Thr Pro Arg

15 ATA CAC ATA CTA GGG AAA GTT CAT AAC TTG CTA GAA ATT
Ile His Ile Leu Gly Lys Val His Asn Leu Leu Glu Ile

ATA CAG GTA GAT ACA CTC ATT CCT ATG AAC AAC ACG CAT
Ile Gln Val Asp Thr Leu Ile Pro MET Asn Asn Thr His

20 ACA AAA GAT GAG GTT AAC AGT TAC CTC ATA CCA CTA AAT
Thr Lys Asp Glu Val Asn Ser Tyr Leu Ile Pro Leu Asn

GCA AAC AGG CAA AAT GAG CAG GTT TTT GGG ACA AAC CTG
Ala Asn Arg Gln Asn Glu Gln Val Phe Gly Thr Asn Leu

25

TTT ATT GGT GAT GGG GTC TTC AAA ACT ACT CTT CTG GGT
Phe Ile Gly Asp Gly Val Phe Lys Thr Thr Leu Leu Gly

30 GAA ATT GTT CAG TAC TAT ACA CAT TGG TCT GGA TCA CTT
Glu Ile Val Gln Tyr Tyr Thr His Trp Ser Gly Ser Leu

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AGA TTC TCT TCG ATG TAT ACT GGT CCT GCC TTG TCC AGT
Arg Phe Ser Ser MET Tyr Thr Gly Pro Ala Leu Ser Ser

5 GCT AAA CTC ACT CTA GCA TAC ACC CCG CCT GGT GCT CGT
Ala Lys Leu Thr Leu Ala Tyr Thr Pro Pro Gly Ala Arg

GGT CCA CAG GAC AGG AGA GAA GCA ATG CTA GGT ACT CAT
Gly Pro Gln Asp Arg Arg Glu Ala MET Leu Gly Thr His

10 GTT GTC TGG GAT ATT GGT CTG CAA TCC ACC ATA GTA ATG
Val Val Trp Asp Ile Gly Leu Gln Ser Thr Ile Val MET

ACA ATA CCA TGG ACA TCA GGG GTG CAG TTT AGA TAT ACT
Thr Ile Pro Trp Thr Ser Gly Val Gln Phe Arg Tyr Thr

15 GAT CCA GAT ACA TAC ACC AGT GCT GGC TTT CTA TCA TGT
Asp Pro Asp Thr Tyr Thr Ser Ala Gly Phe Leu Ser Cys

TGG TAT CAA ACT TCT CTT ATA CTT CCC CCA GAA ACG ACC
20 Trp Tyr Gln Thr Ser Leu Ile Leu Pro Pro Glu Thr Thr

GGC CAG GTC TAC TTA TTA TCA TTC ATA AGT GCA TGT CCA
Gly Gln Val Tyr Leu Leu Ser Phe Ile Ser Ala Cys Pro

25 GAT TTT AAG CTT AGG CTG ATG AAA GAT ACT CAA ACT ATC
Asp Phe Lys Leu Arg Leu MET Lys Asp Thr Gln Thr Ile

VP3 VP1

30 TCA CAG ACT GTT GCA CTC ACT GAA GGC TTA GGT GAT GAA
Ser Gln Thr Val Ala Leu Thr Glu Gly Leu Gly Asp Glu

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TTA GAA GAA GTC ATC GTT GAG AAA ACG AAA CAG ACG GTG
Leu Glu Glu Val Ile Val Glu Lys Thr Lys Gln Thr Val

5 GCC TCA ATC TCA TCT GGT CCA AAA CAC ACA CAA AAA GTC
Ala Ser Ile Ser Ser Gly Pro Lys His Thr Gln Lys Val

CCC ATA CTA ACT GCA AAC GAA ACA GGG GCC ACA ATG CCT
Pro Ile Leu Thr Ala Asn Glu Thr Gly Ala Thr MET Pro

10 GTT CTT CCA TCA GAC AGC ATA GAA ACC AGA ACT ACC TAC
Val Leu Pro Ser Asp Ser Ile Glu Thr Arg Thr Thr Tyr

ATG CAC TTT AAT GGT TCA GAA ACT GAT GTA GAA TGC TTT
MET His Phe Asn Gly Ser Glu Thr Asp Val Glu Cys Phe

15 TTG GGT GGT GCG GCT TGT GTG CAT GTA ACT GAA ATA CAA
Leu Gly Gly Ala Ala Cys Val His Val Thr Glu Ile Gln

AAC AAA GAT GCT ACT GGA ATA GAT AAT CAC AGA GAA GCA
20 Asn Lys Asp Ala Thr Gly Ile Asp Asn His Arg Glu Ala

AGA TTG TTC AAT GAT TGG AAA ATC AAC CTG TCC AGC CTT
Arg Leu Phe Asn Asp Trp Lys Ile Asn Leu Ser Ser Leu

25 GTC CAA CTT AGA AAG AAA CTG GAA CTC TTC ACT TAT GTT
Val Gln Leu Arg Lys Lys Leu Glu Leu Phe Thr Tyr Val

AGG TTT GAT TCT GAG TAT ACC ATA CTG GCC ACT GCA TCT
Arg Phe Asp Ser Glu Tyr Thr Ile Leu Ala Thr Ala Ser

30 CAA CCT GAT TCA GCA AAC TAT TCA AGC AAT TTG GTG GTC
Gln Pro Asp Ser Ala Asn Tyr Ser Ser Asn Leu Val Val

CAA GCC ATG TAT GTT CCA CAT GGT GCC CCG AAA TCC AAA
Gln Ala MET Tyr Val Pro His Gly Ala Pro Lys Ser Lys

5 AGA GTG GAC GAT TAC ACA TGG CAA AGT GCT TCA AAC CCC
Arg Val Asp Asp Tyr Thr Trp Gln Ser Ala Ser Asn Pro

AGT GTA TTC TTC AAG GTG GGG GAT ACA TCA AGG TTT AGT
Ser Val Phe Phe Lys Val Gly Asp Thr Ser Arg Phe Ser

10 GTG CCT TAT GTA GGA TTG GCA TCA GCA TAT AAT TGT TTT
Val Pro Tyr Val Gly Leu Ala Ser Ala Tyr Asn Cys Phe

TAT GAT GGT TAC TCA CAT GAT GAT GCA GAA ACT CAG TAT
15 Tyr Asp Gly Tyr Ser His Asp Asp Ala Glu Thr Gln Tyr

GGC ATA ACT GTT CTA AAC CAT ATG GGT AGT ATG GCA TTC
Gly Ile Thr Val Leu Asn His MET Gly Ser MET Ala Phe

20 AGA ATA GTA AAT GAA CAT GAT GAA CAC TTA ACT CTT GTC
Arg Ile Val Asn Glu His Asp Glu His Leu Thr Leu Val

AAG ATC AGA GTT TAT CAC AGG GCA AAG CTC GTT GAA GCA
Lys Ile Arg Val Tyr His Arg Ala Lys Leu Val Glu Ala

25 TGG ACT CCC AGA GCA CCC AGA GCA CTA CCC TAC ACA TCA
Trp Thr Pro Arg Ala Pro Arg Ala Leu Pro Tyr Thr Ser

ATA GGG CGC ACA AAT TAT CCT AAG AAT ACA GAA CCA GTA
30 Ile Gly Arg Thr Asn Tyr Pro Lys Asn Thr Glu Pro Val

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VP1 3B

ATT AAG AAG AGG AAA GGT GAC ATT AAA TCC TAT GGT TTA
Ile Lys Lys Arg Lys Gly Asp Ile Lys Ser Tyr Gly Leu

5 GGA CCT AGG TAC GGT GGG ATT TAT ACA TCA AAT GTT AAA
Gly Pro Arg Tyr Gly Gly Ile Tyr Thr Ser Asn Val Lys

ATA ATG AAT TAC CAC TTG ATG ACA CCA GAA GAC CAC CAT
Ile MET Asn Tyr His Leu MET Thr Pro Glu Asp His His

10 AAT CTG ATA GCA CCC TAT CCA AAT AGA GAT TTA GCA ATA
Asn Leu Ile Ala Pro Tyr Pro Asn Arg Asp Leu Ala Ile

GTC TCA ACA GGA GGA CAT GGT GCA GAA ACA ATA CCA CAC
15 Val Ser Thr Gly Gly His Gly Ala Glu Thr Ile Pro His

TGT AAC CGT ACA TCA GGT GTT TAC TAT TCC ACA TAT TAC
Cys Asn Arg Thr Ser Gly Val Tyr Tyr Ser Thr Tyr Tyr

20 AGA AAG TAT TAC CCC ATA ATT TGC GAA AGC CCG CCA ACA
Arg Lys Tyr Tyr Pro Ile Ile Cys Glu Ser Pro Pro Thr

TCT GAA TTG GGG AAG CCC TTA TTA CCC AAG CAG ATT CAA
Ser Glu Leu Gly Lys Pro Leu Leu Pro Lys Gln Ile Gln

25 GCA GGA GTG ATG AAA GGG GTT GGG CCG GCA GAG CTA GGA
Ala Gly Val MET Lys Gly Val Gly Pro Ala Glu Leu Gly

GAC TGC GGT GGG ATT TTG AGA TGC ATA CAT GGT CCC ATT
30 Asp Cys Gly Gly Ile Leu Arg Cys Ile His Gly Pro Ile

GGA TTG TTA ACA GCT GAA GGT AGT GGA TAT GTT TGT TTT
Gly Leu Leu Thr Ala Glu Gly Ser Gly Tyr Val Cys Phe

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3B

GCT GAC ATA CGA CAG TTG GAG TGT ATC GCA GAG GAA CAG
Ala Asp Ile Arg Gln Leu Glu Cys Ile Ala Glu Glu Gln

5

5B

GGG CTG AGT GAT TAC ATC ACA GGT TTG GGT AGA GCT TTT
Gly Leu Ser Asp Tyr Ile Thr Gly Leu Gly Arg Ala Phe

10

GGT GTC GGG TTC ACT GAC CAA ATC TCA ACA AAA GTC ACA
Gly Val Gly Phe Thr Asp Gln Ile Ser Thr Lys Val Thr

GAA CTA CAA GAA GTG GCG AAA GAT TTC CTC ACC ACA AAA
Glu Leu Gln Glu Val Ala Lys Asp Phe Leu Thr Thr Lys

15

GTT TTG TCC AAA GTG GTC AAA ATG GTT TCA GCT TTA GTG
Val Leu Ser Lys Val Val Lys MET Val Ser Ala Leu Val

ATC ATT TGC AGA AAT CAT GAT GAC TTG GTC ACT GTT ACG
Ile Ile Cys Arg Asn His Asp Asp Leu Val Thr Val Thr

20

GCC ACT CTA GCA CTA CTT GGA TGT GAT GGA TCT CCT TGG
Ala Thr Leu Ala Leu Leu Gly Cys Asp Gly Ser Pro Trp

25

AGA TTT CTG AAG ATG TAC ATT TCC AAA CAC TTT CAG GTG
Arg Phe Leu Lys MET Tyr Ile Ser Lys His Phe Gln Val

5B X

CCT TAC ATT GAA AGA CAA GCA AAT GAT GGA TGG TTC AGA
Pro Tyr Ile Glu Arg Gln Ala Asn Asp Gly Trp Phe Arg

30

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AAG TTT AAT GAT GCA TGT AAT GCT GCA AAG GGA TTG GAA
Lys Phe Asn Asp Ala Cys Asn Ala Ala Lys Gly Leu Glu

5 TGG ATT GCT AAT AAG ATT TCC AAA CTG ATT GAA TGG ATA
Trp Ile Ala Asn Lys Ile Ser Lys Leu Ile Glu Trp Ile

AAA AAC AAA GTA CTT CCC CAA GCC AAA GAA AAA CTA GAA
Lys Asn Lys Val Leu Pro Gln Ala Lys Glu Lys Leu Glu

10 TTT TGT AGT AAA CTC AAA CAA CTT GAT ATA CTA GAG AGA
Phe Cys Ser Lys Leu Lys Gln Leu Asp Ile Leu Glu Arg

CAA ATA ACC ACC ATG CAT ATC TCG AAT CCA ACA CAG GAA
Gln Ile Thr Thr MET His Ile Ser Asn Pro Thr Gln Glu

15 AAA CGA GAG CAG TTG TTC AAT AAC GTA TTG TGG TTG GAA
Lys Arg Glu Gln Leu Phe Asn Asn Val Leu Trp Leu Glu

CAA ATG TCG CAA AAG TTT GCC CCA TTT TAT GCC GTT GAA
20 Gln MET Ser Gln Lys Phe Ala Pro Phe Tyr Ala Val Glu

TCA AAA AGA ATC AGG GAA CTC AAG AAC AAA ATG GTA AAT
Ser Lys Arg Ile Arg Glu Leu Lys Asn Lys MET Val Asn

25 TAT ATG CAA TTT AAA AGT AAA CAA AGA ACT GAA CCA GTG
Tyr MET Gln Phe Lys Ser Lys Gln Arg Thr Glu Pro Val

TGT GTA TTA ATC CAT GGT ACA CCC GGT TCT GGT AAA TCA
Cys Val Leu Ile His Gly Thr Pro Gly Ser Gly Lys Ser

30 TTA ACA ACA TCC ATT GTG GGA CGT GCA ATT GCA GAA CAC
Leu Thr Thr Ser Ile Val Gly Arg Ala Ile Ala Glu His

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TTC AAT TCA GCA GTA TAT TCA CTT CCA CCA GAT CCC AAG
Phe Asn Ser Ala Val Tyr Ser Leu Pro Pro Asp Pro Lys

5 CAC TTT GAT GGT TAT CAG CAA CAG GAA GTT GTG ATT ATG
His Phe Asp Gly Tyr Gln Gln Gln Glu Val Val Ile MET

GAT GAT CTG AAC CAA AAT CCA GAT GGA CAG GAT ATA AGC
Asp Asp Leu Asn Gln Asn Pro Asp Gly Gln Asp Ile Ser

10 ATG TTT TGT CAA ATG GTT TCT TCA GTG GAT TTC TTG CCT
MET Phe Cys Gln MET Val Ser Ser Val Asp Phe Leu Pro

CCA ATG GCT AGT TTA GAT AAC AAG GGC ATG TTA TTC ACC
15 Pro MET Ala Ser Leu Asp Asn Lys Gly MET Leu Phe Thr

AGT AAT TTT GTT CTA GCC TCC ACA AAT TCT AAC ACA CTA
Ser Asn Phe Val Leu Ala Ser Thr Asn Ser Asn Thr Leu

20 AGC CCC CCA ACA ATC TTG AAT CCT GAA GCT TTA GTC AGG
Ser Pro Pro Thr Ile Leu Asn Pro Glu Ala Leu Val Arg

AGA TTT GGT TTT GAC CTA GAT ATA TGT TTG CAT ACT ACC
Arg Phe Gly Phe Asp Leu Asp Ile Cys Leu His Thr Thr

25 TAC ACA AAG AAT GGA AAA CTC AAT GCA GGC ATG TCA ACC
Tyr Thr Lys Asn Gly Lys Leu Asn Ala Gly MET Ser Thr

AAG ACA TGC AAA GAT TGC CAT CAA CCA TCT AAT TTC AAG
30 Lys Thr Cys Lys Asp Cys His Gln Pro Ser Asn Phe Lys

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AAA TGT TGC CCC CTA GTC TGT GGA AAA GCT ATT AGC TTG
Lys Cys Cys Pro Leu Val Cys Gly Lys Ala Ile Ser Leu

5 GTA GAC AGA ACT ACC AAC GTT AGG TAT AGT GTG GAT CAA
Val Asp Arg Thr Thr Asn Val Arg Tyr Ser Val Asp Gln

CTG GTC ACG GCT ATT ATA AGT GAT TTC AAG AGC AAA ATG
Leu Val Thr Ala Ile Ile Ser Asp Phe Lys Ser Lys MET

10

X 1B

CAA ATT ACA GAT TCC CTA GAA ACA CTG TTT CAA GGA CCA
Gln Ile Thr Asp Ser Leu Glu Thr Leu Phe Gln Gly Pro

15 GTG TAT AAA GAT TTA GAG ATT GAT GTT TGC AAC ACA CCA
Val Tyr Lys Asp Leu Glu Ile Asp Val Cys Asn Thr Pro

CCT TCA GAA TGT ATC AAC GAT TTA CTG AAA TCT GTA GAT
Pro Ser Glu Cys Ile Asn Asp Leu Leu Lys Ser Val Asp

20 TCA GAA GAG ATT AGG GAA TAT TGT AAG AAG AAG AAA TGG
Ser Glu Glu Ile Arg Glu Tyr Cys Lys Lys Lys Lys Trp

ATT ATA CCT GAA ATT CCT ACC AAC ATA GAA AGG GCT ATG
Ile Ile Pro Glu Ile Pro Thr Asn Ile Glu Arg Ala MET

25

AAT CAA GCC AGC ATT ATT ATT AAT ACT ATT CTG ATG TTT
Asn Gln Ala Ser Ile Ile Ile Asn Thr Ile Leu MET Phe

30 GTC AGT ACA TTA GGT ATT GTT TAT GTC ATT TAT AAA TTG
Val Ser Thr Leu Gly Ile Val Tyr Val Ile Tyr Lys Leu

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1B VPg

TTT GCT CAA ACT CAA GGA CCA TAT TCT GGT AAC CCG CCT
Phe Ala Gln Thr Gln Gly Pro Tyr Ser Gly Asn Pro Pro

5 CAC AAT AAA CTA AAA GCC CCA ACT TTA CGC CCA GTT GTT
His Asn Lys Leu Lys Ala Pro Thr Leu Arg Pro Val Val

VPg Protease

GTG CAA GGA CCA AAC ACA GAA TTT GCA CTA TCC CTG TTA
10 Val Gln Gly Pro Asn Thr Glu Phe Ala Leu Ser Leu Leu

AGG AAA AAC ATA ATG ACT ATA ACA ACC TCA AAG GGA GAG
Arg Lys Asn Ile MET Thr Ile Thr Thr Ser Lys Gly Glu

15 TTC ACA GGG TTA GGC ATA CAT GAT CGT GTC TGT GTG ATA
Phe Thr Gly Leu Gly Ile His Asp Arg Val Cys Val Ile

CCC ACA CAC GCA CAG CCT GGT GAT GAT GTA CTA GTG AAT
Pro Thr His Ala Gln Pro Gly Asp Asp Val Leu Val Asn

20 GGT CAG AAA ATT AGA GTT AAG GAT AAG TAC AAA TTA GTA
Gly Gln Lys Ile Arg Val Lys Asp Lys Tyr Lys Leu Val

GAT CCA GAG AAC ATT AAT CTA GAG CTT ACA GTG TTG ACT
25 Asp Pro Glu Asn Ile Asn Leu Glu Leu Thr Val Leu Thr

TTA GAT AGA AAT GAA AAA TTC AGA GAT ATC AGG GGA TTT
Leu Asp Arg Asn Glu Lys Phe Arg Asp Ile Arg Gly Phe

30 ATA TCA GAA GAT CTA GAA GGT GTG GAT GCC ACT TTG GTA
Ile Ser Glu Asp Leu Glu Gly Val Asp Ala Thr Leu Val

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GTA CAT TCA AAT AAC TTT ACC AAC ACT ATC TTA GAA GTT
Val His Ser Asn Asn Phe Thr Asn Thr Ile Leu Glu Val

5 GGC CCT GTA ACA ATG GCA GGA CTT ATT AAT TTG AGT AGC
Gly Pro Val Thr MET Ala Gly Leu Ile Asn Leu Ser Ser

ACC CCC ACT AAC AGA ATG ATT CGT TAT GAT TAT GCA ACA
Thr Pro Thr Asn Arg MET Ile Arg Tyr Asp Tyr Ala Thr

10 AAA ACT GGG CAG TGT GGA GGT GTG CTG TGT GCT ACT GGT
Lys Thr Gly Gln Cys Gly Gly Val Leu Cys Ala Thr Gly

15 AAG ATC TTT GGT ATT CAT GTT GGC GGT AAT GGA AGA CAA
Lys Ile Phe Gly Ile His Val Gly Gly Asn Gly Arg Gln

GGA TTT TCA GCT CAA CTT AAA AAA CAA TAT TTT GTA GAG
Gly Phe Ser Ala Gln Leu Lys Lys Gln Tyr Phe Val Glu

Protease Replicase

20 AAA CAA GGC CAA GTA ATA GCT AGA CAT AAG GTT AGG GAG
Lys Gln Gly Gln Val Ile Ala Arg His Lys Val Arg Glu

25 TTT AAC ATA AAT TCA GTC AAC ACG GCA ACT AAG TCA AAA
Phe Asn Ile Asn Ser Val Asn Thr Ala Thr Lys Ser Lys

TTA CAT CCC AGT GTA TTT TAT GAT GTT TTT CCA GGT GAC
Leu His Pro Ser Val Phe Tyr Asp Val Phe Pro Gly Asp

30 AAG GAA CCT GCT GTA TTG AGT GAC AAT GAT CCC AGA CTG
Lys Glu Pro Ala Val Leu Ser Asp Asn Asp Pro Arg Leu

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GAA GTT AAA TTG ACT GAA TCA TTA TTC TCT AAG TAC AAG
Glu Val Lys Leu Thr Glu Ser Leu Phe Ser Lys Tyr Lys

5 GGG AAT GTA AAT ACG GAA CCC ACT GAA AAT ATG CTT GTG
Gly Asn Val Asn Thr Glu Pro Thr Glu Asn MET Leu Val

GCT GTA GAC CAT TAT GCA GGG CAA CTA TTA TCA CTA GAT
Ala Val Asp His Tyr Ala Gly Gln Leu Leu Ser Leu Asp

10 ATC CCC ACT TCT GAA CTT ACA CTA AAA GAA GCA TTA TAT
Ile Pro Thr Ser Glu Leu Thr Leu Lys Glu Ala Leu Tyr

GGA GTA GAT GGA CTA GAA CCT ATA GAT ATT ACA ACC AGT
Gly Val Asp Gly Leu Glu Pro Ile Asp Ile Thr Thr Ser

15 GCA GGA TTT CCC TAT GTG AGT CTT GGG ATC AAA AAG AGA
Ala Gly Phe Pro Tyr Val Ser Leu Gly Ile Lys Lys Arg

GAC ATT CTG AAT AAA GAG ACC CAG GAC ACA GAA AAG ATG
20 Asp Ile Leu Asn Lys Glu Thr Gln Asp Thr Glu Lys MET

AAG TTT TAT CTA GAC AAG TAT GGC ATT GAC TTG CCT CTA
Lys Phe Tyr Leu Asp Lys Tyr Gly Ile Asp Leu Pro Leu

25 GTT ACA TAT ATT AAG GAT GAA TTA AGA AGT GTT GAC AAA
Val Thr Tyr Ile Lys Asp Glu Leu Arg Ser Val Asp Lys

GTC CGA TTA GGG AAA AGT AGA TTA ATT GAA GCC TCC AGT
Val Arg Leu Gly Lys Ser Arg Leu Ile Glu Ala Ser Ser

30 TTG AAT GAT TCT GTT AAC ATG AGA ATG AAA CTA GGC AAC
Leu Asn Asp Ser Val Asn MET Arg MET Lys Leu Gly Asn

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CTT TAC AAA GCA TTC CAT CAA AAT CCC GGT GTT CTG ACT
Leu Tyr Lys Ala Phe His Gln Asn Pro Gly Val Leu Thr

5 GGA TCA GCA GTG GGT TGT GAT CCT GAT GTG TTT TGG TCT
Gly Ser Ala Val Gly Cys Asp Pro Asp Val Phe Trp Ser

GTC ATC CCT TGC TTA ATG GAT GGG CAC CTG ATG GCA TTT
Val Ile Pro Cys Leu MET Asp Gly His Leu MET Ala Phe

10 GAT TAC TCT AAT TTT GAT GCC TCT TTG TCA CCA GTT TGG
Asp Tyr Ser Asn Phe Asp Ala Ser Leu Ser Pro Val Trp

TTT GTC TGT CTA GAG AAG GTT TTG ACC AAG TTA GGC TTT
15 Phe Val Cys Leu Glu Lys Val Leu Thr Lys Leu Gly Phe

GCA GGC TCT TCA TTA ATT CAA TCA ATT TGT AAT ACC CAT
Ala Gly Ser Ser Leu Ile Gln Ser Ile Cys Asn Thr His

20 CAT ATC TTT AGG GAT GAA ATA TAT GTG GTT GAA GGT GGC
His Ile Phe Arg Asp Glu Ile Tyr Val Val Glu Gly Gly

ATG CCC TCA GGG TGT TCA GGA ACC AGC ATA TTC AAT TCC
MET Pro Ser Gly Cys Ser Gly Thr Ser Ile Phe Asn Ser

25 ATG ATC AAC AAC ATA ATC ATT AGG ACT TTG ATA TTA GAT
MET Ile Asn Asn Ile Ile Ile Arg Thr Leu Ile Leu Asp

GCA TAT AAA GGA ATA GAT TTA GAC AAA CTT AAA ATC TTA
30 Ala Tyr Lys Gly Ile Asp Leu Asp Lys Leu Lys Ile Leu

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GCT TAC GGT GAT GAT TTG ATT GTT TCT TAT CCT TAT GAA
Ala Tyr Gly Asp Asp Leu Ile Val Ser Tyr Pro Tyr Glu

CTG GAT CCA CAA GTG TTG GCA ACT CTT GGT AAA AAT TAT
5 Leu Asp Pro Gln Val Leu Ala Thr Leu Gly Lys Asn Tyr

GGA CTA ACC ATC ACA CCC CCA GAC AAA TCT GAA ACT TTT
Gly Leu Thr Ile Thr Pro Pro Asp Lys Ser Glu Thr Phe

10 ACA AAA ATG ACA TGG GAA AAC TTG ACA TTT TTA AAG AGA
Thr Lys MET Thr Trp Glu Asn Leu Thr Phe Leu Lys Arg

TAC TTC AAG CCT GAT CAA CAA TTT CCC TTT TTG GTT CAC
Tyr Phe Lys Pro Asp Gln Gln Phe Pro Phe Leu Val His

15 CCA GTT ATG CCC ATG AAA GAT ATA CAT GAG TCA ATC AGA
Pro Val MET Pro MET Lys Asp Ile His Glu Ser Ile Arg

TGG ACA AAG GAT CCT AAA AAC ACA CAG GAT CAC GTC CGA
20 Trp Thr Lys Asp Pro Lys Asn Thr Gln Asp His Val Arg

TCA TTA TGC ATG TTA GCA TGG CAC TCA GGA GAA AAA GAG
Ser Leu Cys MET Leu Ala Trp His Ser Gly Glu Lys Glu

25 TAC AAT GAA TTC ATT CAG AAG ATC AGA ACT ACT GAC ATT
Tyr Asn Glu Phe Ile Gln Lys Ile Arg Thr Thr Asp Ile

GGA AAA TGT CTA ATT CTC CCA GAA TAC AGC GTA CTT AGG
Gly Lys Cys Leu Ile Leu Pro Glu Tyr Ser Val Leu Arg

30

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Replicase

AGG CGC TGG TTG GAC CTC TTT

Arg Arg Trp Leu Asp Leu Phe

5 TAGGTTAACAATATAGACACTTAATTTGAGTAGAAGTAGGAGT

TTATAAAAAA AAAAAAAA

2. DNA representative of genomic RNA of VP4
10 of HRV Type 14 having the nucleotide and
corresponding amino acid sequence:

VP4

ATG GGC GCT CAG GTT TCT ACA CAG AAA AGT GGA TCT CAC
MET Gly Ala Gln Val Ser Thr Gln Lys Ser Gly Ser His

15

GAA AAT CAA AAC ATT TTG ACC AAT GGA TCA AAT CAG ACT
Glu Asn Gln Asn Ile Leu Thr Asn Gly Ser Asn Gln Thr

20

TTC ACA GTT ATA AAT TAC TAT AAG GAT GCA GCA AGT ACA
Phe Thr Val Ile Asn Tyr Tyr Lys Asp Ala Ala Ser ThrTCA TCA GCT GGT CAA TCA CTG TCA ATG GAC CCA TCT AAG
Ser Ser Ala Gly Gln Ser Leu Ser MET Asp Pro Ser Lys

25

TTT ACA GAA CCA GTT AAA GAT CTC ATG CTT AAG GGT GCA
Phe Thr Glu Pro Val Lys Asp Leu MET Leu Lys Gly Ala

VP4

CCA GCA TTG ATT

30

Pro Ala Leu Asn

3. DNA representative of genomic RNA of VP2
of HRV Type 14 having the nucleotide and
corresponding amino acid sequence:

VP2

5 TCA CCC AAT GTT GAG GCC TGT GGT TAT
Ser Pro Asn Val Glu Ala Cys Gly Tyr

AGT GAT AGA GTA CAA CAA ATC ACA CTC GGG AAT TCA ACA
Ser Asp Arg Val Gln Gln Ile Thr Leu Gly Asn Ser Thr

10

ATA ACA ACA CAA GAA GCA GCC AAC GCT GTT GTG TGT TAT
Ile Thr Thr Gln Glu Ala Ala Asn Ala Val Val Cys Tyr

15 GCT GAA TGG CCA GAG TAC CTT CCA GAT GTG GAC GCT AGT
Ala Glu Trp Pro Glu Tyr Leu Pro Asp Val Asp Ala Ser

GAT GTC AAT AAA ACT TCA AAA CCA GAC ACT TCT GTC TGT
Asp Val Asn Lys Thr Ser Lys Pro Asp Thr Ser Val Cys

20 AGG TTT TAC ACA TTG GAT AGT AAG ACA TGG ACA ACA GGT
Arg Phe Tyr Thr Leu Asp Ser Lys Thr Trp Thr Thr Gly

TCT AAA GGC TGG TGC TGG AAA TTA CCA GAT GCA CTC AAG
Ser Lys Gly Trp Cys Trp Lys Leu Pro Asp Ala Leu Lys

25

GAT ATG GGT GTG TTC GGG CAA AAC ATG TTT TTC TAC TCA
Asp MET Gly Val Phe Gly Gln Asn MET Phe Phe Tyr Ser

30 CTA GGA AGA TCA GGT TAC ACA GTA CAC GTT CAG TGC AAT
Leu Gly Arg Ser Gly Tyr Thr Val His Val Gln Cys Asn

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171121A

GCC ACA AAA TTC CAT AGC GGT TGT CTA CTT GTA GTT GTA
Ala Thr Lys Phe His Ser Gly Cys Leu Leu Val Val Val

5 ATA CCA GAA CAC CAA CTG GCT TCA CAT GAG GGT GGC AAT
Ile Pro Glu His Gln Leu Ala Ser His Glu Gly Gly Asn

GTT TCA GTT AAA TAC ACA TTC ACG CAT CCA GGT GAA CGT
Val Ser Val Lys Tyr Thr Phe Thr His Pro Gly Glu Arg

10 GGT ATA GAT TTA TCA TCT GCA AAT GAA GTG GGA GGG CCT
Gly Ile Asp Leu Ser Ser Ala Asn Glu Val Gly Gly Pro

GTC AAG GAT GTC ATA TAC AAT ATG AAT GGT ACT TTA TTA
Val Lys Asp Val Ile Tyr Asn MET Asn Gly Thr Leu Leu
15

GGA AAT CTG CTC ATT TTC CCT CAC CAG TTC ATT AAT CTA
Gly Asn Leu Leu Ile Phe Pro His Gln Phe Ile Asn Leu

20 AGA ACC AAT AAT ACA GCC ACA ATA GTG ATA CCA TAC ATA
Arg Thr Asn Asn Thr Ala Thr Ile Val Ile Pro Tyr Ile

AAC TCA GTA CCC ATT GAT TCA ATG ACA CGT CAC AAC AAT
Asn Ser Val Pro Ile Asp Ser MET Thr Arg His Asn Asn

25 GTC TCA CTG ATG GTC ATC CCT ATT GCC CCT CTT ACA GTA
Val Ser Leu MET Val Ile Pro Ile Ala Pro Leu Thr Val

CCA ACT GGA GCA ACT CCC TCA CTC CCT ATA ACA GTC ACA
Pro Thr Gly Ala Thr Pro Ser Leu Pro Ile Thr Val Thr
30

ATA GCA CCT ATG TGC ACT GAG TTC TCT GGG ATA AGG TCC
Ile Ala Pro MET Cys Thr Glu Phe Ser Gly Ile Arg Ser

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VP2

AAG TCA ATT GTG CCA CAA
Lys Ser Ile Val Pro Gln

5

4. DNA representative of genomic RNA of VP1
of HRV type 14 having the nucleotide and
corresponding amino acid sequence:

VP1

10 GGC TTA GGT GAT GAA
Gly Leu Gly Asp Glu

TTA GAA GAA GTC ATC GTT GAG AAA ACG AAA CAG ACG GTG
Leu Glu Glu Val Ile Val Glu Lys Thr Lys Gln Thr Val

15

GCC TCA ATC TCA TCT GGT CCA AAA CAC ACA CAA AAA GTC
Ala Ser Ile Ser Ser Gly Pro Lys His Thr Gln Lys Val

20 CCC ATA CTA ACT GCA AAC GAA ACA GGG GCC ACA ATG CCT
Pro Ile Leu Thr Ala Asn Glu Thr Gly Ala Thr MET Pro

GTT CTT CCA TCA GAC AGC ATA GAA ACC AGA ACT ACC TAC
Val Leu Pro Ser Asp Ser Ile Glu Thr Arg Thr Thr Tyr

25 ATG CAC TTT AAT GGT TCA GAA ACT GAT GTA GAA TGC TTT
MET His Phe Asn Gly Ser Glu Thr Asp Val Glu Cys Phe

TTG GGT GGT GCG GCT TGT GTG CAT GTA ACT GAA ATA CAA
Leu Gly Gly Ala Ala Cys Val His Val Thr Glu Ile Gln

30

AAC AAA GAT GCT ACT GGA ATA GAT AAT CAC AGA GAA GCA
Asn Lys Asp Ala Thr Gly Ile Asp Asn His Arg Glu Ala

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AGA TTG TTC AAT GAT TGG AAA ATC AAC CTG TCC AGC CTT
Arg Leu Phe Asn Asp Trp Lys Ile Asn Leu Ser Ser Leu

5 GTC CAA CTT AGA AAG AAA CTG GAA CTC TTC ACT TAT GTT
Val Gln Leu Arg Lys Lys Leu Glu Leu Phe Thr Tyr Val

AGG TTT GAT TCT GAG TAT ACC ATA CTG GCC ACT GCA TCT
Arg Phe Asp Ser Glu Tyr Thr Ile Leu Ala Thr Ala Ser

10 CAA CCT GAT TCA GCA AAC TAT TCA AGC AAT TTG GTG GTC
Gln Pro Asp Ser Ala Asn Tyr Ser Ser Asn Leu Val Val

CAA GCC ATG TAT GTT CCA CAT GGT GCC CCG AAA TCC AAA
15 Gln Ala MET Tyr Val Pro His Gly Ala Pro Lys Ser Lys

AGA GTG GAC GAT TAC ACA TGG CAA AGT GCT TCA AAC CCC
Arg Val Asp Asp Tyr Thr Trp Gln Ser Ala Ser Asn Pro

20 AGT GTA TTC TTC AAG GTG GGG GAT ACA TCA AGG TTT AGT
Ser Val Phe Phe Lys Val Gly Asp Thr Ser Arg Phe Ser

GTG CCT TAT GTA GGA TTG GCA TCA GCA TAT AAT TGT TTT
Val Pro Tyr Val Gly Leu Ala Ser Ala Tyr Asn Cys Phe

25 TAT GAT GGT TAC TCA CAT GAT GAT GCA GAA ACT CAG TAT
Tyr Asp Gly Tyr Ser His Asp Asp Ala Glu Thr Gln Tyr

GGC ATA ACT GTT CTA AAC CAT ATG GGT AGT ATG GCA TTC
30 Gly Ile Thr Val Leu Asn His MET Gly Ser MET Ala Phe

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AGA ATA GTA AAT GAA CAT GAT GAA CAC TTA ACT CTT GTC
Arg Ile Val Asn Glu His Asp Glu His Leu Thr Leu Val

AAG ATC AGA GTT TAT CAC AGG GCA AAG CTC GTT GAA GCA
5 Lys Ile Arg Val Tyr His Arg Ala Lys Leu Val Glu Ala

TGG ACT CCC AGA GCA CCC AGA GCA CTA CCC TAC ACA TCA
Trp Thr Pro Arg Ala Pro Arg Ala Leu Pro Tyr Thr Ser

10 ATA GGG CGC ACA AAT TAT CCT AAG AAT ACA GAA CCA GTA
Ile Gly Arg Thr Asn Tyr Pro Lys Asn Thr Glu Pro Val

VP1

ATT AAG AAG AGG AAA GGT GAC ATT AAA TCC TAT
15 Ile Lys Lys Arg Lys Gly Asp Ile Lys Ser Tyr

5. DNA representative of genomic RNA of
protease of HRV type 14 having the nucleotide and
corresponding amino acid sequence:

20 Protease
GGA CCA AAC ACA GAA TTT GCA CTA TCC CTG TTA
Gly Pro Asn Thr Glu Phe Ala Leu Ser Leu Leu

AGG AAA AAC ATA ATG ACT ATA ACA ACC TCA AAG GGA GAG
25 Arg Lys Asn Ile MET Thr Ile Thr Thr Ser Lys Gly Glu

TTC ACA GGG TTA GGC ATA CAT GAT CGT GTC TGT GTG ATA
Phe Thr Gly Leu Gly Ile His Asp Arg Val Cys Val Ile

30 CCC ACA CAC GCA CAG CCT GGT GAT GAT GTA CTA GTG AAT
Pro Thr His Ala Gln Pro Gly Asp Asp Val Leu Val Asn

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GGT CAG AAA ATT AGA GTT AAG GAT AAG TAC AAA TTA GTA
Gly Gln Lys Ile Arg Val Lys Asp Lys Tyr Lys Leu Val

5 GAT CCA GAG AAC ATT AAT CTA GAG CTT ACA GTG TTG ACT
Asp Pro Glu Asn Ile Asn Leu Glu Leu Thr Val Leu Thr

TTA GAT AGA AAT GAA AAA TTC AGA GAT ATC AGG GGA TTT
Leu Asp Arg Asn Glu Lys Phe Arg Asp Ile Arg Gly Phe

10 ATA TCA GAA GAT CTA GAA GGT GTG GAT GCC ACT TTG GTA
Ile Ser Glu Asp Leu Glu Gly Val Asp Ala Thr Leu Val

GTA CAT TCA AAT AAC TTT ACC AAC ACT ATC TTA GAA GTT
Val His Ser Asn Asn Phe Thr Asn Thr Ile Leu Glu Val

15 GGC CCT GTA ACA ATG GCA GGA CTT ATT AAT TTG AGT AGC
Gly Pro Val Thr MET Ala Gly Leu Ile Asn Leu Ser Ser

ACC CCC ACT AAC AGA ATG ATT CGT TAT GAT TAT GCA ACA
20 Thr Pro Thr Asn Arg MET Ile Arg Tyr Asp Tyr Ala Thr

AAA ACT GGG CAG TGT GGA GGT GTG CTG TGT GCT ACT GGT
Lys Thr Gly Gln Cys Gly Gly Val Leu Cys Ala Thr Gly

25 AAG ATC TTT GGT ATT CAT GTT GGC GGT AAT GGA AGA CAA
Lys Ile Phe Gly Ile His Val Gly Gly Asn Gly Arg Gln

GGA TTT TCA GCT CAA CTT AAA AAA CAA TAT TTT GTA GAG
Gly Phe Ser Ala Gln Leu Lys Lys Gln Tyr Phe Val Glu

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Protease

AAA CAA

Lys Gln

- 5 6. DNA representative of genomic RNA of
replicase of HRV type 14 having the nucleotide and
corresponding amino acid sequence:

Replicase

- GGC CAA GTA ATA GCT AGA CAT AAG GTT AGG GAG
10 Gly Gln Val Ile Ala Arg His Lys Val Arg Glu
- TTT AAC ATA AAT TCA GTC AAC ACG GCA ACT AAG TCA AAA
Phe Asn Ile Asn Ser Val Asn Thr Ala Thr Lys Ser Lys
- 15 TTA CAT CCC AGT GTA TTT TAT GAT GTT TTT CCA GGT GAC
Leu His Pro Ser Val Phe Tyr Asp Val Phe Pro Gly Asp
- AAG GAA CCT GCT GTA TTG AGT GAC AAT GAT CCC AGA CTG
Lys Glu Pro Ala Val Leu Ser Asp Asn Asp Pro Arg Leu
- 20 GAA GTT AAA TTG ACT GAA TCA TTA TTC TCT AAG TAC AAG
Glu Val Lys Leu Thr Glu Ser Leu Phe Ser Lys Tyr Lys
- GGG AAT GTA AAT ACG GAA CCC ACT GAA AAT ATG CTT GTG
25 Gly Asn Val Asn Thr Glu Pro Thr Glu Asn MET Leu Val
- GCT GTA GAC CAT TAT GCA GGG CAA CTA TTA TCA CTA GAT
Ala Val Asp His Tyr Ala Gly Gln Leu Leu Ser Leu Asp
- 30 ATC CCC ACT TCT GAA CTT ACA CTA AAA GAA GCA TTA TAT
Ile Pro Thr Ser Glu Leu Thr Leu Lys Glu Ala Leu Tyr

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GGA GTA GAT GGA CTA GAA CCT ATA GAT ATT ACA ACC AGT
Gly Val Asp Gly Leu Glu Pro Ile Asp Ile Thr Thr Ser

5 GCA GGA TTT CCC TAT GTG AGT CTT GGG ATC AAA AAG AGA
Ala Gly Phe Pro Tyr Val Ser Leu Gly Ile Lys Lys Arg

GAC ATT CTG AAT AAA GAG ACC CAG GAC ACA GAA AAG ATG
Asp Ile Leu Asn Lys Glu Thr Gln Asp Thr Glu Lys MET

10 AAG TTT TAT CTA GAC AAG TAT GGC ATT GAC TTG CCT CTA
Lys Phe Tyr Leu Asp Lys Tyr Gly Ile Asp Leu Pro Leu

GTT ACA TAT ATT AAG GAT GAA TTA AGA AGT GTT GAC AAA
Val Thr Tyr Ile Lys Asp Glu Leu Arg Ser Val Asp Lys

15 GTC CGA TTA GGG AAA AGT AGA TTA ATT GAA GCC TCC AGT
Val Arg Leu Gly Lys Ser Arg Leu Ile Glu Ala Ser Ser

TTG AAT GAT TCT GTT AAC ATG AGA ATG AAA CTA GGC AAC
20 Leu Asn Asp Ser Val Asn MET Arg MET Lys Leu Gly Asn

CTT TAC AAA GCA TTC CAT CAA AAT CCC GGT GTT CTG ACT
Leu Tyr Lys Ala Phe His Gln Asn Pro Gly Val Leu Thr

25 GGA TCA GCA GTG GGT TGT GAT CCT GAT GTG TTT TGG TCT
Gly Ser Ala Val Gly Cys Asp Pro Asp Val Phe Trp Ser

GTC ATC CCT TGC TTA ATG GAT GGG CAC CTG ATG GCA TTT
Val Ile Pro Cys Leu MET Asp Gly His Leu MET Ala Phe

30 GAT TAC TCT AAT TTT GAT GCC TCT TTG TCA CCA GTT TGG
Asp Tyr Ser Asn Phe Asp Ala Ser Leu Ser Pro Val Trp

TTT GTC TGT CTA GAG AAG GTT TTG ACC AAG TTA GGC TTT
Phe Val Cys Leu Glu Lys Val Leu Thr Lys Leu Gly Phe

5 GCA GGC TCT TCA TTA ATT CAA TCA ATT TGT AAT ACC CAT
Ala Gly Ser Ser Leu Ile Gln Ser Ile Cys Asn Thr His

CAT ATC TTT AGG GAT GAA ATA TAT GTG GTT GAA GGT GGC
His Ile Phe Arg Asp Glu Ile Tyr Val Val Glu Gly Gly

10 ATG CCC TCA GGG TGT TCA GGA ACC AGC ATA TTC AAT TCC
MET Pro Ser Gly Cys Ser Gly Thr Ser Ile Phe Asn Ser

ATG ATC AAC AAC ATA ATC ATT AGG ACT TTG ATA TTA GAT
15 MET Ile Asn Asn Ile Ile Ile Arg Thr Leu Ile Leu Asp

GCA TAT AAA GGA ATA GAT TTA GAC AAA CTT AAA ATC TTA
Ala Tyr Lys Gly Ile Asp Leu Asp Lys Leu Lys Ile Leu

20 GCT TAC GGT GAT GAT TTG ATT GTT TCT TAT CCT TAT GAA
Ala Tyr Gly Asp Asp Leu Ile Val Ser Tyr Pro Tyr Glu

CTG GAT CCA CAA GTG TTG GCA ACT CTT GGT AAA AAT TAT
Leu Asp Pro Gln Val Leu Ala Thr Leu Gly Lys Asn Tyr

25 GGA CTA ACC ATC ACA CCC CCA GAC AAA TCT GAA ACT TTT
Gly Leu Thr Ile Thr Pro Pro Asp Lys Ser Glu Thr Phe

ACA AAA ATG ACA TGG GAA AAC TTG ACA TTT TTA AAG AGA
30 Thr Lys MET Thr Trp Glu Asn Leu Thr Phe Leu Lys Arg

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TAC TTC AAG CCT GAT CAA CAA TTT CCC TTT TTG GTT CAC
Tyr Phe Lys Pro Asp Gln Gln Phe Pro Phe Leu Val His

CCA GTT ATG CCC ATG AAA GAT ATA CAT GAG TCA ATC AGA
5 Pro Val MET Pro MET Lys Asp Ile His Glu Ser Ile Arg

TGG ACA AAG GAT CCT AAA AAC ACA CAG GAT CAC GTC CGA
Trp Thr Lys Asp Pro Lys Asn Thr Gln Asp His Val Arg

10 TCA TTA TGC ATG TTA GCA TGG CAC TCA GGA GAA AAA GAG
Ser Leu Cys MET Leu Ala Trp His Ser Gly Glu Lys Glu

TAC AAT GAA TTC ATT CAG AAG ATC AGA ACT ACT GAC ATT
Tyr Asn Glu Phe Ile Gln Lys Ile Arg Thr Thr Asp Ile

15 GGA AAA TGT CTA ATT CTC CCA GAA TAC AGC GTA CTT AGG
Gly Lys Cys Leu Ile Leu Pro Glu Tyr Ser Val Leu Arg

Replicase

20 AGG CGC TGG TTG GAC CTC TTT
Arg Arg Trp Leu Asp Leu Phe

7. A hybridoma cell made by fusing a
mammalian myeloma cell to a spleen cell from a mammal
25 immunized with a serotype of the major group of human
rhinoviruses of HRV or a protein subunit thereof.

8. A hybridoma cell according to Claim 7
wherein the protein subunit is VP1, VP2 or VP3.

30 9. A monoclonal antibody to HRV produced by
a hybridoma cell of Claim 7.

10. A monoclonal antibody produced by a hybridoma cell made by fusion of a mammalian myeloma cell to a spleen cell from a mammal immunized with a) cells containing receptor sites for serotypes of the major group of human rhinoviruses or with b) cell membranes containing receptor sites for serotypes of the major group of human rhinoviruses or with c) cells and cell membranes containing receptor sites for serotypes of the major group of human rhinoviruses.

11. A monoclonal antibody of Claim 10 wherein the spleen cell is from a mammal immunized with cells containing receptor sites for serotypes of the major group of human rhinoviruses.

12. A monoclonal antibody of Claim 10 wherein the spleen cell is from a mammal immunized with cell membranes containing receptor sites for serotypes of the major group of human rhinoviruses.

13. A monoclonal antibody of Claim 10 wherein the spleen cell is from a mammal immunized with cells and cell membranes containing receptor sites for serotypes of the major group of human rhinoviruses.

14. An antibody according to Claim 10 wherein the mammalian myeloma cell is from a rodent.

15. An antibody according to Claim 14
wherein the rodent is a mouse.

5 16. An antibody according to Claim 10 which
prevents attachment of serotypes of the major group
of human rhinoviruses to a susceptible mammalian cell.

10 17. An antibody according to Claim 16
wherein the antibody prevents attachment for up to
about 12 hours.

15 18. An antibody according to Claim 10 which
is capable of displacing at least some of the
serotypes of the major group of human rhinoviruses
when the serotypes are bound to a susceptible
mammalian cell.

20 19. A Fab fragment produced by enzyme
digestion of the antibody of Claim 10.

25 20. An in vitro composition comprising a
continuous hybridoma cell line which secretes the
antibody of Claim 10.

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